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410 Rec'd PCT/PTO 12 MAY 2000

FORM-PTO-1390 (Rev. 5-93)		U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE	ATTORNEY'S DOCKET NUMBER 2488-1-001
TRANSMITTAL LETTER TO THE UNITED STATES DESIGNATED/ELECTED OFFICE (DO/EO/US) CONCERNING A FILING UNDER 35 U.S.C. 371			U.S. APPLICATION NO. (If known, see 37 C.F.R. 1.5) 09/554547
INTERNATIONAL APPLICATION NO. PCT/GB98/03397	INTERNATIONAL FILING DATE 12 November 1998	PRIORITY DATE CLAIMED 12 November 1997	
TITLE OF INVENTION TISSUE CEMENT PROTEINS FROM RHIPICEPHALUS APPENDICULATUS			
APPLICANT(S) FOR DO/EO/US Guido Christian PAESEN; Patricia Ann NUTTALL			
Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:			
1. <input checked="" type="checkbox"/> This is a FIRST submission of items concerning a filing under 35 U.S.C. 371. 2. <input type="checkbox"/> This is a SECOND or SUBSEQUENT submission of items concerning a filing under 35 U.S.C. 371. 3. <input checked="" type="checkbox"/> This express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and the PCT Articles 22 and 39(1). 4. <input checked="" type="checkbox"/> A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date. 5. <input checked="" type="checkbox"/> A copy of the International Application as filed (35 U.S.C. 371(c)(2)) a. <input checked="" type="checkbox"/> is transmitted herewith (required only if not transmitted by the International Bureau). b. <input checked="" type="checkbox"/> has been transmitted by the International Bureau. c. <input type="checkbox"/> is not required, as the application was filed in the United States Receiving Office (RO/US). 6. <input checked="" type="checkbox"/> A translation of the International Application into English (35 U.S.C. 371(c)(2)). 7. <input checked="" type="checkbox"/> Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3)) a. <input checked="" type="checkbox"/> are transmitted herewith (required only if not transmitted by the International Bureau). b. <input type="checkbox"/> have been transmitted by the International Bureau. c. <input type="checkbox"/> have not been made; however, the time limit for making such amendments has NOT expired. d. <input type="checkbox"/> have not been made and will not be made. 8. <input type="checkbox"/> A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)). 9. <input checked="" type="checkbox"/> An unexecuted oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)). 10. <input type="checkbox"/> A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)). Items 11. to 16. below concern other document(s) or information included: 11. <input type="checkbox"/> An Information Disclosure Statement under 37 CFR 1.97 and 1.98. 12. <input type="checkbox"/> An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included. 13. <input checked="" type="checkbox"/> A FIRST preliminary amendment. <input type="checkbox"/> A SECOND or SUBSEQUENT preliminary amendment. 14. <input type="checkbox"/> A substitute specification. 15. <input type="checkbox"/> A change of power of attorney and/or address letter. 16. <input checked="" type="checkbox"/> Other items or information: NINETEEN (19) SHEETS OF DRAWINGS; INTERNATIONAL SEARCH REPORT; WRITTEN OPINION; INTERNATIONAL PRELIMINARY EXAMINATION REPORT; NOTIFICATION OF RECORDING OF CHANGE			
EXPRESS MAIL "MAILING CERTIFICATE NO.": EL485953902US DATE OF DEPOSIT: MAY 12, 2000			

U.S. APPLICATION NO. (If known, see 37 C.F.R. § 501.103) 09/554547		INTERNATIONAL APPLICATION NO. PCT/GB98/03397		ATTORNEY'S DOCKET NUMBER 2488-1-001	
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17. <input checked="" type="checkbox"/> The following fees are submitted:	CALCULATIONS	PTO USE ONLY
Basic National Fee (37 CFR 1.492(a)(1)-(5)): Search Report has been prepared by the EPO or JPO \$840.00 International preliminary examination fee paid to USPTO (37 CFR 1.482) \$670.00 No international preliminary examination fee paid to USPTO (37 CFR 1.482) but international search fee paid to USPTO (37 CFR 1.445(a)(2)) \$760.00 Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$970.00 International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(2)-(4) \$ 96.00 <div style="text-align: right;">ENTER APPROPRIATE BASIC FEE AMOUNT =</div>		
Surcharge of \$130.00 for furnishing the oath or declaration later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(e)).		
Claims	Number Filed	Number Extra
Total Claims	56 -20 =	36
Independent Claims	1 -3 =	0
Multiple dependent claim(s) (if applicable)		+ \$260.00
TOTAL OF ABOVE CALCULATIONS =		\$ 1,748.00
Reduction for 1/2 for filing by small entity, if applicable. Verified Small Entity statement must also be filed. (Note 37 CFR 1.9, 1.27, 1.28).		\$
SUBTOTAL =		\$ 1,748.00
Processing fee of \$130.00 for furnishing the English translation later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(f)).		\$
TOTAL NATIONAL FEE =		\$ 1,748.00
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property +		\$
TOTAL FEES ENCLOSED =		\$
		Amount to be: refunded \$
		charged \$

a. ☒ A check in the amount of \$ 1,748.00 to cover the above fees is enclosed.

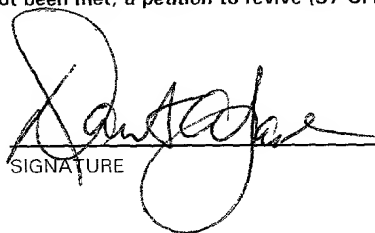
b. ☐ Please charge my Deposit Account No. 11-1153 in the amount of \$ _____ to cover the above fees. A duplicate copy of this sheet is enclosed.

c. ☒ The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. 11-1153. A duplicate copy of this sheet is enclosed.

NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

SEND ALL CORRESPONDENCE TO:

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 SIGNATURE
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David A. Jackson
 REGISTRATION NUMBER 26,742

EXPRESS MAIL "MAILING CERTIFICATE NO.": EL485953902US DATE OF DEPOSIT: MAY 12, 2000

09/554547
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PATENT
2488-1-001

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

APPLICANTS : Guido Christian PAESEN; Patricia Ann NUTTALL
APPLICATION NO. : PCT/GB98/03397
FILED : 12 November 1998
FOR : TISSUE CEMENT PROTEINS FROM RHIPICEPHALUS
APPENDICULATUS

PRELIMINARY AMENDMENT

ASSISTANT COMMISSIONER FOR PATENTS
BOX PCT
WASHINGTON, D.C. 20231

Sir:

Prior to calculating the fees pursuant to the entry into the National Phase of the above-identified Application, please amend the claims as follows:

IN THE CLAIMS:

Claim 4, line 1, delete "any one of the preceding claims" and insert --Claim 1--.
Claim 5, line 1, delete "any one of the preceding claims" and insert --Claim 1--.
Claim 9, line 1, delete "any one of the preceding claims" and insert --Claim 1--.
Claim 11, line 1, delete "any one of the preceding claims" and insert --Claim 1--.
Claim 12, line 1, delete "any one of the preceding claims" and insert --Claim 1--.
Claim 15, line 1, delete "any one of the preceding claims" and insert --Claim 1--.
Claim 16, line 1, delete "any one of the preceding claims" and insert --Claim 1--.
Claim 17, line 1, delete "any one of the preceding claims" and insert --Claim 1--.
Claim 18, line 1, delete "any one of the preceding claims" and insert --Claim 1--.

Claim 19, line 1, delete "any one of the preceding claims" and insert --Claim 1--.

Claim 20, line 2, delete "any preceding";
line 2, after "claim" insert --1--.

Claim 21, line 2, delete "any one of claims 1 to 19" and insert --Claim 1--.

Claim 22, line 2, delete "any one of claims 1 to 19" and insert --Claim 1--.

Please amend Claim 23 as follows:

Claim 23. (Amended) The tissue cement protein according to [any one of] claim[s] 1 [to 19] or a pharmaceutical composition of any one of claims 20 [to] 21 or 22 for use in therapy.

Claim 24, line 1, delete "any one of claims 1 to 19" and insert --Claim 1--.

Claim 25, line 1, delete "any one of claims 1 to 19" and insert --Claim 1--.

Claim 26, line 1, delete "any one of claims 1 to 19" and insert --Claim 1--.

Claim 27, lines 1 and 2, delete "any one of claims 1 to 19" and insert --Claim 1--.

Claim 28, line 2, delete "any one of claims 1 to 19" and insert --Claim 1--.

Claim 29, lines 2 and 3, delete "any one of claims 1 to 19" and insert --Claim 1--.

Please amend Claim 30 as follows:

Claim 30. (Amended) The tissue cement portion of [any one of] claim[s] 1 [to 19] or a pharmaceutical composition of any one of claims 20 [to] or 22 for use in the temporary or permanent bonding of tissues.

Claim 31, line 1, delete "any one of claims 1 to 19" and insert --Claim 1--.

Claim 32, lines 1 and 2, delete "any one of claims 1 to 19" and insert --Claim 1--.

Claim 35, line 2, delete "any one of claims 32 to 34" and insert --Claim 32--.

Claim 38, line 2, delete "to" and insert --,36 or--.

Please amend Claims 39 and 40 as follows:

39. (Amended) A transgenic animal that has been transformed by the nucleic acid molecule of [any one of] claim[s] 31 to 34] 32 or a vector of one of claims 35 [to 38] , 36 or 37.

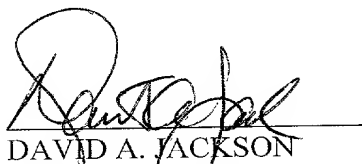
40. (Amended) A method of preparing a tissue cement protein of [any one of] claim[s] 1 [to 19] comprising expressing a vector according to any one of claims 35 [to] , 36 or 37 in a host cell and culturing said host cell under conditions where said protein is expressed, and recovering said protein thus expressed.

REMARKS

The above amendments are submitted herewith to reduce multiple dependencies and to conform the claims more closely to U.S. practice.

Entry of the foregoing amendments and early and favorable processing in the National Phase before the United States Patent and Trademark Office is courteously solicited.

Respectfully submitted,



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Applicant or Patentee: Guido Christian ^a PAESEN: Patricia Anne NUTTALL
Application or Patent No.: 09/554,547
Filed or Issued: May 12, 2000
For: TISSUE CEMENT

**VERIFIED STATEMENT (DECLARATION) CLAIMING SMALL ENTITY
STATUS (37 C.F.R. §§ 1.9(f) AND 1.27(d)) - NONPROFIT ORGANIZATION**

I hereby declare that I am an official empowered to act on behalf of the nonprofit organization identified below:

NAME OF ORGANIZATION THE NATURAL ENVIRONMENTAL RESEARCH COUNCIL
ADDRESS OF ORGANIZATION POLARIS HOUSE, NORTH STAR AVENUE, SWINDON,
SN2 1EU, UNITED KINGDOM

TYPE OF ORGANIZATION

- ☒ University or other institution of higher education
☐ Tax exempt under Internal Revenue Service Code (26 U.S.C. §§ 501(a) and 501(c)(3))
☐ Nonprofit scientific or educational under statute of state of The United States of America
(Name of state _____)
(Citation of statute _____)
☐ Would qualify as tax exempt under Internal Revenue Service Code (26 U.S.C. §§ 501(a) and 501(c)(3)) if located in The United States of America
☐ Would qualify as nonprofit scientific or educational under statute of The United States of America if located in The United States of America
(Name of state _____)
(Citation of statute _____)

I hereby declare that the nonprofit organization identified above qualifies as a nonprofit organization as defined in 37 C.F.R. § 1.9(e) for purposes of paying reduced fees under Sections 41(a) and 41(b) of Title 35, United States Code, with regard to the invention entitled by inventor(s) PAESEN G C: NUTTALL P A described in

- ☐ the specification filed herewith
☒ Application No. 09/554,547, filed May 12, 2000
☐ Patent No. _____, issued _____

I hereby declare that rights under contract or law have been conveyed to and remain with the nonprofit organization with regard to the above-identified invention.

If the rights held by the above-identified nonprofit organization are not exclusive, each individual, concern, or organization having rights to the invention is listed below,* and no rights to the invention are held by any person, other than the inventor, who would not qualify as an individual inventor under 37 C.F.R. § 1.9(c), or by any concern that would not qualify as either a small business concern under 37 C.F.R. § 1.9(d) or a nonprofit organization under 37 C.F.R. § 1.9(e).

*NOTE: Separate verified statements are required from each named person, concern, or organization having rights to the invention averring to their status as small entities. (37 C.F.R. § 1.27.)

FULL NAME _____

ADDRESS _____

☐ individual ☐ small business concern ☐ nonprofit organization

FULL NAME _____

ADDRESS _____

☐ individual ☐ small business concern ☐ nonprofit organization

I acknowledge the duty to file, in this application or patent, notification of any change in status resulting in loss of entitlement to small entity status prior to paying, or at the time of paying, the earlier of the issue fee and any maintenance fee due after the date on which status as a small entity is no longer appropriate. (37 C.F.R. § 1.28(b).)

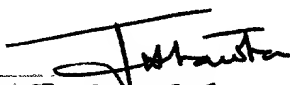
I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code; and that such willful false statements may jeopardize the validity of the application, any patent issuing thereon, or any patent to which this verified statement is directed.


NAME OF PERSON SIGNING JOHN LAWTON (CHIEF EXECUTIVE) AND JOHN HANSFORD (DIRECTOR OF PERSONNEL)

TITLE IN ORGANIZATION _____

ADDRESS OF PERSON SIGNING NERC, POLARIS HOUSE, SWINDON, WILTS,
ENGLAND, UK

SIGNATURE _____ DATE 7/3/00


CHIEF EXECUTIVE


DIRECTOR, PERSONNEL & ESTATES

ON BEHALF OF THE NATURAL ENVIRONMENT
RESEARCH COUNCIL

POLARIS HOUSE
NORTH STAR AVENUE
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WILTSHIRE, ENGLAND

TISSUE CEMENT PROTEINS FROM RHIPICEPHALUS APPENDICULATUS

The present invention relates to tissue cement proteins produced by certain species of blood-feeding ectoparasites. These proteins and compositions comprising these 5 proteins are particularly useful for the temporary or permanent bonding of animal tissues to each other or to other biomaterials. The present invention also relates to the use of tissue cement proteins in the production of vaccines that protect animals against the bite of blood-sucking ectoparasites and the transmission of viruses, bacteria and other pathogens by such ectoparasites.

10

Cement is produced by many blood-feeding ectoparasites, including certain species of Ixodid ticks. Ixodid (hard) ticks are haematophagous parasites that attach themselves to a vertebrate host by means of a 'cement cone', a product of the type II and type III acini of the tick salivary glands (Kemp *et al.*, 1982; Walker *et al.*, 1985). (All 15 documents referred to herein are listed at the end of the description.)

The cement that forms the cone is a milky-white secretion that is injected into the skin of animals on which these parasites feed. The cement comprises a number of interacting protein and carbohydrate components. The cement spreads into the bite site 20 and over the skin and, upon hardening, ensures that the mouthparts remain firmly anchored to the host during the feeding period, which typically lasts 4 to 8 days. The cement cone functions additionally as a gasket to prevent leakage of fluids from the bite site during feeding.

25 The tick cement cone is a layered structure, constructed from two major types of cement. The first type of cement is produced just minutes after establishing the bite site and hardens quickly to form a rigid 'core' of the cone. A second type of cement is secreted later, about 24 hours after attachment, and hardens more slowly to form a more flexible 'cortex'. In adult ticks, cement production typically continues until the 30 3rd or 4th day after attachment (Kemp *et al.*, 1982; Sonenshine *et al.*, 1991).

The tick cement cone appears to be mainly proteinaceous, but also contains some carbohydrate and lipid. An early study found the amino-acid composition of whole cement in *Boophilus microplus* to be rich in glycine, leucine, serine and tyrosine (Kemp *et al.*, 1982). However, the individual proteins comprising the tick cement are
5 very poorly characterised. Although the mobility of the component proteins has been shown on SDS-PAGE gels, none have yet been purified.

The process by which the cement components harden is also not understood, although mechanisms similar to the tanning of cuticle and coagulation of haemolymph have
10 been proposed (Kemp *et al.*, 1982; Moorhouse and Tatchell, 1966). At present no direct scientific evidence has been produced to substantiate these theoretical mechanisms.

It has been noted that the polypeptides that form the cement cortex appear to be similar
15 to certain structural components of vertebrate skin. Involvement of these vertebrate-like molecules may enable ticks to use host-derived enzymes during the cement hardening process, for example: lysyl oxidases which cross-link collagen and elastin (Siegel, 1979); or transglutaminases, such as the coagulation factor XIIIa, which is induced during wound healing and cross-links fibronectin, fibrins and collagen
20 (Ichinose *et al.*, 1990). These enzymes may cross-link cortex polypeptides to the extracellular matrix proteins of the skin.

Other enzymes such as phenoloxidases or peroxidases which catalyse the hardening of arthropod extracellular structures (Sugumaran *et al.*, 1992) have been identified in *R.*
25 *appendiculatus* salivary glands and are therefore likely to play a role in solidifying the cement cone.

The composition of tick cement appears to be similar amongst different Ixodid tick species. For example, an antiserum raised against a 90kD salivary protein of the
30 brown ear tick, *Rhipicephalus appendiculatus*, has been shown to recognise polypeptides from the salivary glands and cement proteins of the American dog tick, *Dermacentor variabilis*, the lone star tick, *Amblyomma americanum*, and the brown

dog tick, *R. sanguineus* (Jaworski *et al.*, 1992).

All these tick species are extremely effective as transmitters of disease. For example,
5 *R. appendiculatus* represents a major obstacle to livestock development in several sub-
saharan regions. It transmits the protozoan parasite *Theileria parva* which causes the
usually fatal East Coast Fever. This disease is often considered the most important
disease of cattle (Norval *et al.*, 1992a; Norval *et al.*, 1992b). This tick is also the main
vector of the virus causing Nairobi sheep disease, a disabling and often deadly disease
10 in sheep and goats (Davies, 1988). *R. appendiculatus* and other tick pests also cause
considerable damage to the skin, thereby affecting the leather industry.

In an effort to combat parasite-transmitted diseases, unpurified cement components
have been tested as inducers of host resistance (Brown *et al.*, 1986; Shapiro *et al.*,
15 1989), but reliable vaccines based on cement proteins have not been successfully
developed. Cone proteins would appear to be a reasonable target for a vaccine since
the formation of the cone is essential for the tick to attach to the host and feed.
However, only some of the cone proteins are antigenic.

20 There therefore exists a great need for an effective vaccine to combat diseases that are
transmitted by blood-feeding ectoparasites. The elucidation of the components of
tissue cement produced by these organisms would allow the rational design of such
vaccines.

25 Furthermore, these molecules would prove useful in medicine as components of tissue
cement. Presently available tissue cements are of two types, both of which suffer from
significant disadvantages. Acrylic-based glues are extremely strong, yet are also very
toxic and can thus only be used in very small quantities in the body. The second type
of tissue cement used is non-immunogenic but forms a much less strong bond.
30 Consequently this type of cement is only useful in a small number of surgical
procedures. There is thus a great need for a non-immunogenic tissue cement that is
capable of bonding mammalian tissue with great strength.

Summary of the invention

According to the present invention there is provided a tissue cement protein having the amino acid sequence shown in Figure 3 or Figure 7 or containing any one of the partial amino acid sequences shown in any one of Figures 2, 4 to 6 and 8, related tissue cement proteins from blood-feeding parasites, preferably ticks, and functional equivalents thereof.

The proteins of the present invention are of two subtypes: group A and group B. Proteins in group A form the cement cone core when secreted in saliva and harden quickly to form a rigid latex-like structure. The proteins in group B form the cortex of the cement cone and harden more slowly. The resulting structure is more flexible. Accordingly, functional equivalents of proteins of either group A or group B will possess these respective activities and properties.

The term "functional equivalents" is used herein to describe those proteins that have an analogous function to tissue cement proteins containing the amino acid sequences identified in any one of Figures 2 to 8.

These proteins may belong to the same protein family as the proteins and partial proteins identified in Figures 2 to 8. By protein family is meant a group of polypeptides that share a common function and exhibit common sequence homology between motifs present in the polypeptide sequences.

By sequence homology is meant that the polypeptide sequences are related by divergence from a common ancestor. In particular, as is discussed in more detail below, the proteins and partial proteins identified herein possess certain sequences in common that are repeated several times throughout the sequence of the protein. Preferably, the homology between polypeptide sequences is at least 50% across the whole of the amino acid sequence of the protein. More preferably, the homology is at least 75% across the whole of the amino acid sequence of the protein. Most preferably, homology is greater than 80% across the whole of the protein sequence.

By "analogous function" is meant firstly that the proteins have retained the capacity to form a cement. Such proteins will thus be capable of hardening over a period of time to form a solid mass or glue. Secondly, this term may refer to proteins that are structurally similar to group A or group B proteins and thus contain similar or identical epitopes. These functional equivalents may thus be used as immunogens to develop vaccines, directed against blood-feeding parasites, that target members of the tissue cement protein family.

- 10 Functional equivalents of tissue cement proteins may include, for example, mutants containing amino acid substitutions, insertions or deletions from the wild type sequence. Functional equivalents with improved function from that of the wild type sequence may also be designed through the systematic or directed mutation of specific residues in the protein sequence. Improvements in function that may be desired will include greater strength of bonding, faster speed of bonding or greater flexibility of the hardened cement.

Functional equivalents of tissue cement proteins or protein fragments may be made more or less immunogenic than the corresponding wild type protein or protein fragment in order to suit a desired application. If the proteins are to be used in surgical procedures as tissue cements then the proteins should ideally be non-immunogenic to evade attack by the immune system. However, if the tissue cement proteins are to be used in a vaccination regime to induce host resistance to parasite proteins, then the proteins may be modified so as to enhance their immunogenicity. They will thus be more likely to elicit an immune response in the vaccinated host.

Functional equivalents will include conservative amino acid substitutions that do not affect the function or activity of the protein in an adverse manner. This term is also intended to include natural biological variants (e.g. allelic variants or geographical variations within the species from which the tissue cement proteins are derived).

According to the invention, fragments of tissue cement proteins are also envisioned as functional equivalents. Fragments that have retained those portions of the protein that are responsible for a desired activity may prove ideal in certain applications. For example, short stretches of peptide derived from immunogenic portions of tissue
5 cement proteins will be useful as immunogens. An antibody will normally recognise an epitope comprising between six and twelve amino acid residues. Such short stretches of polypeptide sequence are simple to produce in large quantities, either synthetically or through recombinant means.

- 10 The tissue cement proteins of the present invention may function either as a structural component of tissue cement or may possess an enzymatic activity directed against the structural components of the tissue cement.

It is thought that most of the protein and partial protein sequences so far identified and
15 shown in Figures 2 to 8 are structural components of tissue cement. The applicant, however, does not wish to be bound by this theory. For example, the protein sequence identified in Figure 2 appears to contain a signal sequence and its sequence resembles that of keratin, a widely studied structural protein. Similarly, the protein whose sequence is set out in Figure 3 also contains a signal sequence and is glycine and
20 proline rich, like many structural proteins. The cemA protein, whose partial sequence is illustrated in Figure 4, contains a number of repeats and is thus also likely to be a structural component of tissue cement.

The protein of Figure 5 is composed of a number of repeats and resembles collagen in
25 sequence. The encoding cDNA shares sequences in common with glutenin, a known self-assembling protein. It thus seems likely that this protein is capable of self-assembly. The applicant, however, does not wish to be bound by this theory. The possibility that this particular sequence may be involved in self-assembly raises the opportunity of using these motifs to bestow on an unrelated protein the ability to self-
30 assemble.

In common with some of the other proteins illustrated in the accompanying Figures, the protein of Figure 6 contains a number of consensus recognition sites for carbohydrate moieties, in particular glycosaminoglycans.

5 The protein sequence illustrated in Figure 7 also contains consensus attachment sites for glycosaminoglycan moieties and possesses a putative signal sequence. The amino terminal half of the protein resembles collagen, whilst the carboxy terminal shares more in common with keratin. The protein is glycine-rich and contains several repeats of the motif (C/S)1-4(Y/F) which is also found in structural proteins from the egg
10 shells of certain insects. The tyrosines in these consensus sequences may be involved in the cross-linking of this protein through the formation of dityrosine bridges by the action of phenoloxidases.

The sequence of Figure 8 is both glycine and tyrosine rich and resembles a cement
15 protein of the reef-building polychaete *Pragmatopoma californica* (see Figure 9). It is thus likely that this protein is also a structural component of tissue cement. The applicant, however, does not wish to be bound by this theory.

The enzymatic activity that may be possessed by the tissue cement proteins of the
20 present invention may involve the ability to effect such covalent modifications as phosphorylation, glycosylation, reduction or oxidation of other proteins and carbohydrate moieties and may result in the cross-linking of the structural components of the tissue cement. Cross-linking may be either reversible or irreversible and may occur between homologous or heterologous components of the tissue cement. The
25 cross-linking may also occur between tissue cement proteins and non-parasite proteins such as, for example, components of vertebrate tissue.

The tissue cement proteins of the present invention may be group A proteins. By "group A" is meant that in parasite saliva these proteins form the core of the cement
30 cone. The function of the cone core in parasites is to attach to the skin of a vertebrate host and to form a rigid bond that will not break easily. Accordingly, these proteins form a hard latex-like cement that sets and bonds to the vertebrate skin quickly. Group

A proteins are thus ideally suited to applications that require a quick-setting tough bond.

The tissue cement proteins of the present invention may be group B proteins. By
5 "group B" is meant that in tick saliva these proteins form the cortex of the cement cone. One function of the cortex in parasites is to form a gasket-like seal around the bite site, to prevent leakage of fluids. A further function of the cortex proteins is to form a flexible hinge so that the parasite will not be easily brushed off its host. Accordingly, group B proteins harden more slowly than group A proteins, but set to
10 form a more flexible, pliant cement. These proteins are thus ideally suited for applications when a more flexible bond is required.

Many of the structural tissue cement proteins of the present invention share in common the ability to bind to vertebrate tissue. This binding may be due to an inherent
15 affinity possessed by the protein for certain components of the vertebrate skin, such as collagen. However, an affinity for vertebrate proteins may only manifest itself when in the presence of enzymes whose activity is required in order to generate an association between a tissue cement protein and a component of the skin. This enzymatic activity may be derived from tissue cement proteins themselves or may be provided by enzyme
20 components of the vertebrate skin, such as lysyl oxidases, that cross-link collagen and elastin or transglutaminases, such as the coagulation factor XIIIa, that cross-link fibronectin, fibrins and collagen during many vertebrate healing processes.

The tissue cement proteins or their functional equivalents according to the present
25 invention may be derived from any blood-feeding parasite. Preferably, the tissue cement proteins of the present invention are derived from blood-feeding ectoparasites, more preferably ticks. Most preferably, the tissue cement proteins of the present invention are derived from the brown ear tick *Rhipicephalus appendiculatus*.

30 The tissue cement proteins of the present invention may also comprise carbohydrate components. Many tissue cement proteins are in fact glycoproteins, containing carbohydrate attachments covalently bound at various sites in the protein. The

carbohydrate generally will comprise a series of monosaccharide units that commonly occur as an oligosaccharide or fairly small polysaccharide. As has been discussed briefly above, many of the proteins so far identified in ticks possess consensus attachment sites for glycosaminoglycans (glycans containing aminosaccharide 5 residues).

The tissue cement proteins of the invention may be present in the tissue cement as monomers, dimers, tetramers, or as oligomers comprising a number of homologous or heterologous monomers as one unit. This is particularly true of structural tissue cement 10 proteins, which may associate non-covalently as part of the cement hardening process. The applicant, however, does not wish to be bound by this theory.

The tissue cement proteins of the present invention may be purified from cement produced by live parasites. This may be done by treating collected cones with a wash 15 solution such as PBS, a TRIS buffer or non-ionic detergents, for example Tween-20 or Triton. Cement proteins may be prepared through immunoprecipitation using antibodies that are specific for epitopes in the protein sequence. Alternatively, the proteins may be prepared synthetically, or using techniques of genetic engineering. Preferably, the tissue cement proteins of the present invention comprise recombinant 20 polypeptides produced by expression from an encoding nucleic acid.

Synthetic molecules designed to mimic the tertiary structure or active site of the tissue cement proteins constitute a further aspect of the invention.

25 A further aspect of the present invention comprises tissue cement proteins that are fused to other molecules such as labels, toxins or bioactive molecules. Particularly suitable candidates for fusion will be reporter molecules such as luciferase, green fluorescent protein or horse radish peroxidase. Linker molecules such as streptavidin or biotin may also be used. Additionally, bioactive peptides or polypeptides may be 30 fused to a tissue cement protein. Such molecules may comprise molecules with antiseptic or antibiotic properties, or toxins for targeting to cancer cells.

The proteins may be fused chemically, using methods such as chemical cross-linking. Such methods will be well known to those of skill in the art and may comprise, for example, cross-linking of the thiol groups of cysteine residues. Chemical cross-linking will in most instances be used to fuse tissue cement proteins to non-protein molecules, 5 such as labels. The labels may be radiolabels or labels that can be detected spectroscopically, for example fluorescent or phosphorescent chemical groups.

When it is desired to fuse a tissue cement protein to another protein molecule, the method of choice will often be to fuse the molecules genetically. In order to generate a 10 recombinant fusion protein, the genes or gene portions that encode the proteins or protein fragments of interest are engineered so as to form one contiguous gene arranged so that the codons of the two gene sequences are transcribed in frame.

A tissue cement protein may be fused genetically to any protein for which the 15 encoding gene sequence is or becomes known. Particularly suitable candidates for fusion will be reporter molecules such as luciferase, green fluorescent protein, biotin, avidin, streptavidin or horse radish peroxidase. Additionally, toxin peptides or polypeptides may be fused to a tissue cement protein. Antiseptic or antibiotic proteins and peptides may also be fused to the tissue cement proteins of the present invention.

20

According to a further aspect of the present invention there is provided a pharmaceutical composition comprising a tissue cement comprising a mixture of group A and group B tissue cement proteins in the absence of other parasite saliva proteins, optionally in the presence of one or more compounds capable of cross- 25 linking said tissue cement proteins, in conjunction with a pharmaceutically-acceptable excipient.

Such a pharmaceutical composition has many applications, particularly in skin surgery and wound healing, for the temporary or permanent bonding of human or animal 30 tissues to each other or to other biomaterials.

Tissue cement has previously been used in surgical procedures to provide adhesion and stability to living tissues to enable the normal processes of healing and repair to take place or to provide a long term bond in situations where normal healing is delayed or unlikely to occur. Tissue cement formed from the proteins of the present invention have several advantages over conventional tissue cements. For example, the proteins of the invention form strong bonds with vertebrate tissues. This makes them ideal for use as components of a tissue cement to bond two tissue surfaces or edges together.

Different surgical procedures necessitate the use of tissue cement with different properties. Tissue cement formed from the proteins of the present invention is therefore ideal, since the hardening or elastic properties of the cement may be tailored precisely to provide the particular requirements of the surgical procedure through modification of the relative amounts of group A and group B proteins that the cement comprises. The tissue cement is in this manner extremely versatile.

Tissue cement with a high content of group A proteins will generally be useful for procedures that require an extremely tough bond that will not need to flex to any great extent. A good example of such a bond might be a bond between two bone surfaces or between a bone surface and the surface of an artificial joint. A high group A content tissue cement will also be required when it is necessary that the bond sets quickly.

For procedures such as the bonding of skin lacerations, where a high degree of flexibility is required, a tissue cement will be used that contains a high content of group B proteins. However, group B proteins do bond more slowly, so tissue cement with a high group B content will not generally be used for procedures that require a tough bond to form rapidly. However, this cement may be used in conjunction with other measures, such as surgical staples or group A tissue cement, that can form a quick bond to hold the tissues together while the group B proteins harden.

Pharmaceutical compositions that comprise tissue cement proteins according to the present invention may also contain additional preservatives, or components responsible for the prevention of premature setting. The composition may also

comprise a propellant, for instance, if the tissue cement is to be sprayed onto tissue surfaces. Such compounds will be well known to those of skill in the art.

One important advantage of the tissue cement proteins of the present invention is that, 5 *in situ* in vertebrate tissue these proteins are non-immunogenic and therefore do not cause inflammation of the tissue. This is particularly relevant when the tissue cement is intended for internal use, for example in the securing of prolapsed organs. Were the tissue cement immunogenic, an immune attack would be directed against the tissue cement, so causing local inflammation, disrupting the cement and preventing the 10 permanent bonding of the tissue.

The current rationale in surgery to overcome immune rejection involves combatting the initial rejection phenomena until, eventually, the immune system becomes tolerant. For example, after organ transplants, huge combined doses of immunosuppressive 15 agents are initially required that are gradually reduced during treatment until, if the transplant successfully survives for a year or so, very little maintenance immunotherapy is required.

The tissue cement proteins of the present invention can also be used to prevent 20 immune rejection. Aside from their natural non-immunogenicity, another property of the tissue cement proteins of the present invention is that the proteins themselves bind to bioactive proteins in the saliva of an ectoparasite, such as tick histamine-binding proteins. By doing this, the cement proteins localise the action of the various immunosuppressive molecules produced by the organism. These immunosuppressive 25 compounds alter over time during the course of feeding in order to adapt to the host's rejection response. Hence the cement cone, in effect, is an active local immunosuppressive structure.

The ability to bind to bioactive parasite proteins means that tissue cement comprising 30 tissue cement proteins according to the present invention may be complemented with certain immunosuppressive molecules produced by some parasites. For example, suitable molecules might include vasoactive amine binding molecules that are the

subject of co-pending International patent application PCT/GB97/01372. These molecules will bind to the tissue cement, and so provide a local immunosuppressive structure that will disguise any allogeneic or even xenogeneic organ and prevent its rejection.

5

These properties of the tissue cement proteins of the present invention thus provide an addition to the conventional uses of tissue cement to hold articulated joints to bone or to repair damaged organs. Aside from its use as a non-immunogenic adhesive, tissue cement produced according to the present invention may be used as an adjunct to
10 organ transplantation to supply and localise immunosuppressive compounds to the locality of the transplant.

A further aspect of the present invention is therefore to provide tissue cement with immunosuppressive properties by complementation of the basic cement with one or
15 more tissue cement proteins of the present invention, optionally in conjunction with parasite-derived immunosuppressive compounds.

Other specific applications of tissue cement formed from the tissue cement proteins of the present invention may include, but will not be confined to, those given in the
20 following list:

repair of incised surgical wounds in place of conventional closures such as sutures or staples; repair of lacerations such as perineal tears following childbirth or flap lacerations after trauma; the grafting of skin, cultured skin substitutes or biomaterial substitutes to chronic wounds, burns, skin graft donor sites and in reconstructive or
25 cosmetic plastic surgery; the securing of myoplastic flaps; tissue to tissue repair such as repair of lacerations of the liver or other parenchymatous organs; tissue to tissue anastomosis such as gastrointestinal anastomosis or tissue to biomaterial anastomosis such as vascular repair or anastomosis; tissue to tissue reconstruction such as the securing of prolapsed organs in their anatomical site; tissue to tissue approximation as
30 in pleuro-pleural adhesion following recurrent pneumothorax; as a sclerosant in procedures such as the injection of haemorrhoids; in neurosurgical procedures such as the repair or patching of dura mater with natural or artificial substitutes and the

grafting or repair of severed neural tissue; in orthodontic procedures such as the re-implantation of teeth or the reconstruction of the mandibular or maxillary arches using bone chips or materials such as collagen matrices or hydroxyapatite; in orthopaedic procedures including vertebral fusion, arthrodesis, fixation of fractures and
5 osteotomies and the implantation of prostheses such as hip arthroplasty; in procedures such as rhytidectomy where the use of sutures is undesirable on cosmetic grounds; securing of artificial materials such as Marlex™ mesh in hernioplasty (performed in open operation or endoscopically); repair of friable tissue such as tendon or muscle; as a haemostat on oozing surfaces or in circumstances where haemostasis by
10 conventional means is difficult to secure (for example during endoscopic surgery); for the obliteration of sinuses and fistulae; and for sealing perforations of hollow viscera such as the stomach or duodenum (either used alone or in conjunction with a tissue or artificial patch.

15 In a still further embodiment of the invention, the use of tissue cement proteins is provided as tools in the study of cement cone assembly and in the development of strategies to prevent cement cone assembly, and thereby inhibit tick attachment and feeding. For example, monoclonal antibodies and engineered vaccines directed against tissue cement proteins could be used to prevent parasite feeding.

20

Arthropod parasites are sources of infectious disease agents such as tick-borne encephalitis virus, Crimean-Congo haemorrhagic fever virus, Nairobi sheep virus, *Borrelia burgdorferi* (the agent of Lyme disease), *Theileria parva* (the agent of East Coast fever) and other injurious effects that have major impacts in human and
25 veterinary medicine. Control of the arthropod parasites currently relies primarily on the use of chemicals such as acaricides.

Attempts have been made to use immunological means of control through vaccine technology. Some success has been met in identifying certain protective antigens of
30 arthropod parasites as being potential vaccine candidates, but only a few have as yet come to commercial fruition, most notably for the cattle tick *Boophilus microplus*. Despite these developments, there is nonetheless a continuing need for arthropod

parasite vaccines and in particular for a vaccine which may be used against ticks.

An alternative vaccine strategy that has until now not been possible is to vaccinate animals using purified antigens. The immune system of the animal thus develops an improved humoral response to these antigenic polypeptides and correspondingly develops resistance against the arthropod parasites themselves.

One disadvantage of using vaccines directed against a specific vector-transmitted disease to control that disease is that a different vaccine is usually required to protect against each disease. Vaccines directed against disease vectors (such as ticks and mosquitoes) have an added advantage in that their effect may control several different infections, as long as these infections are transmitted by only one type of disease vector.

The present invention therefore also provides for the use of tissue cement proteins as defined above as immunogens. Accordingly, a further aspect of the present invention comprises a vaccine comprising one or more tissue cement proteins as defined above in an arthropod parasite vaccine and in particular as protective immunogens in the control of diseases caused by infections transmitted by arthropod parasites.

20

The vaccine may be administered singly, or in combination with other immunogens. The vaccine may include adjuvants of the type which are well known in the art, for example, alum. Suitable candidates for vaccination include humans and domesticated animals such as cattle, goats, sheep, dogs, cats and other mammalia. All these species require protection against arthropod parasites, particularly ticks, and the infections they transmit.

According to a further aspect of the present invention there is provided a nucleic acid molecule encoding a tissue cement protein as defined above, or any functionally equivalent form. The nucleic acid sequences of choice comprise or contain the nucleic acid sequences exhibited in Figures 2 to 8. The skilled man will appreciate that changes may be made at the nucleotide level by addition, substitution, deletion or

insertion of one or more nucleotides, which changes may or may not be reflected at the amino acid level, dependent on the degeneracy of the genetic code.

The nucleic acid molecule according to this aspect of the present invention may
5 comprise DNA, RNA or cDNA and may additionally comprise nucleotide analogues
in its sequence. Preferably, the nucleic acid comprises DNA, more preferably single or
double-stranded cDNA.

Antisense sequences may also be designed with respect to the nucleic acids of this
10 aspect of the invention and in sequence will in whole or in part comprise that of the
complementary strand to the coding nucleic acid strand. Oligonucleotides comprising
antisense sequences to tissue cement protein genes may be used as diagnostic tools in
the detection of organisms or vectors expressing nucleic acids that encode tissue
cement proteins. These single-stranded oligonucleotides will comprise lengths of
15 nucleic acid of between 10 and 300 nucleotides, preferably between 10 and 100
nucleotides, most preferably of between 10 and 30 nucleotides. The oligonucleotides
may be labelled in order to aid their detection. Suitable labelling systems are well
known in the art.

20 Methods for screening cDNA libraries for proteins analogous to the tissue cement
proteins described herein will be apparent to the man of skill in the art. The antisense
sequences of this aspect of the invention therefore may not correspond exactly to the
complementary strand of the nucleic acid that encodes a tissue cement protein. For
example, when using antisense oligonucleotides as probes in the screening of a cDNA
25 library for proteins analogous to the tissue cement proteins described herein, due to the
degeneracy of the genetic code and inter-species sequence divergence, any analogous
genes to those described herein are likely to comprise sequences that are significantly
different to that of the probe.

30 Accordingly, antisense sequences for use in accordance with this aspect of the present
invention comprise sequences that hybridise under standard conditions to the nucleic
acid sequences exhibited in Figures 2 to 8. 'Hybridising sequences' included within

the scope of the invention are those binding under standard conditions. As used herein, by 'standard conditions' is meant both non-stringent standard hybridisation conditions (6 x SSC/50% formamide at room temperature) with washing under conditions of low stringency (2 x, room temperature, or 2 x SSC, 42°C) or at standard conditions of higher stringency, e.g. 2 x SSC, 65°C (where SSC = 0.15M NaCl, 0.015M sodium citrate, pH 7.2). Preferably standard conditions refers to conditions of high stringency.

A further aspect of the present invention comprises a method of production of a tissue cement protein which method comprises expression from the encoding nucleic acid therefor. Expression may conveniently be achieved by culturing under appropriate conditions recombinant host cells containing the nucleic acid.

Systems for cloning and expression of a polypeptide in a variety of different host cells are well known. Suitable host cells include bacteria, mammalian cells, yeast and baculovirus systems. Mammalian cell lines available in the art for expression of a heterologous polypeptide include Chinese hamster ovary cells, HeLa cells, baby hamster kidney cells and many others. A common, preferred bacterial host is *E. coli*.

The expression of heterologous polypeptides and polypeptide fragments in prokaryotic cells such as *E. coli* is well established in the art; see for example *Molecular Cloning: a Laboratory Manual*: 2nd edition, Sambrook et al., 1989. Cold Spring Harbor Laboratory Press. Expression in eukaryotic cells in culture is also an option available to those skilled in the art for the production of heterologous proteins; see recent reviews, for example O'Reilly *et al.*, (1994), Baculovirus expression vectors - a laboratory manual, Oxford University Press.

Suitable vectors can be chosen or constructed for expression of tissue cement proteins, containing the appropriate regulatory sequences, including promoter sequences, terminator sequences, polyadenylation sequences, enhancer sequences, marker genes and other sequences as appropriate. Vectors may be plasmids, viral e.g. bacteriophage, or phagemid, as appropriate. For further details see *Molecular Cloning: a Laboratory Manual* (loc. cit). Many known techniques and protocols for manipulation of nucleic

acid, for example, in the preparation of nucleic acid constructs. mutagenesis, sequencing, introduction of DNA into cells and gene expression, and analysis of proteins. are described in detail in *Short Protocols in Molecular Biology*, Second Edition, Ausubel et al. eds., John Wiley & Sons, 1992. For example, in eukaryotic
5 cells, the vectors of choice are virus-based, such as baculovirus-based.

A further aspect of the present invention provides a host cell containing a nucleic acid encoding a tissue cement protein or functional equivalent thereof. A still further aspect provides a method comprising introducing such nucleic acid into a host cell or
10 organism. Introduction of nucleic acid may employ any available technique. In eukaryotic cells, suitable techniques may include calcium phosphate transfection, DEAE-Dextran-mediated transfection, electroporation, liposome-mediated transfection or transduction using retrovirus or other viruses, such as vaccinia or, for insect cells, baculovirus. In bacterial cells, suitable techniques may include calcium chloride
15 transformation, electroporation or transfection using bacteriophage.

Introduction of the nucleic acid may be followed by causing or allowing expression from the nucleic acid, e.g. by culturing host cells under conditions for expression of the gene.

20

In one embodiment, the nucleic acid of the invention is integrated into the genome (e.g. chromosome) of the host cell. Integration may be promoted by inclusion of sequences which promote recombination with the genome, in accordance with standard techniques.

25

Transgenic animals transformed so as to express or overexpress in the germ line one or more tissue cement proteins or functional equivalents as described herein form a still further aspect of the invention, along with methods for their production. Many techniques now exist to introduce transgenes into the embryo or germ line of an
30 organism, such as those illustrated in Watson *et al.*, (1994) *Recombinant DNA* (2nd edition), Scientific American Books.

Various aspects and embodiments of the present invention will now be described in more detail by way of example, with particular reference to tissue cement proteins isolated from ticks, and especially from *Rhipicephalus appendiculatus*. It will be appreciated that modification of detail may be made without departing from the scope of the invention.

Brief description of the Figures

Figure 1A is an SDS-gel containing male and female salivary gland extract (SGE) and male and female cement polypeptides obtained by rinsing cement cones in PBS.

Figure 1B is the corresponding Western blot to Figure 1A, probed with a polyclonal antiserum raised against a 17 Kd protein purified from tick salivary gland extract.

Figure 2 is a partial cDNA sequence and translation product of clone 21. The cDNA-inferred protein is a cement protein; it contains a hydrophobic N-terminal region which possibly constitutes a signal sequence, typical for secreted proteins. The protein strongly resembles other structural proteins, especially keratin. A recognition sequence for post-translational attachment of glycosaminoglycan groups is underlined.

Figure 3 is the cDNA and cDNA-inferred polypeptide sequence of clone 33. A putative signal sequence is given in bold. Like many structural proteins, this protein is glycine- and proline-rich. The protein also displays some resemblance to keratins.

Figure 4 is a partial sequence of *cemA* cDNA and the cDNA-inferred polypeptide sequence. The protein is very repetitive, with the sequence KGALLQQQQASQVKGALKAI, or slight variants thereof, repeated several times.

Figure 5 is a partial cDNA and cDNA-inferred polypeptide sequence of clone 24. The protein has resemblance to structural proteins (amongst others collagen), and is contains repeats. The cDNA also has a region in common with glutenin, a self-assembling protein.

Figure 6 is a partial cDNA and cDNA-inferred sequence of clone 68. The encoded proteins resemble structural proteins, such as keratin. A series of putative glycosaminoglycan attachment sites are underlined.

5

Figure 7 is the complete cDNA sequence and cDNA-inferred polypeptide sequence of clone 64. The putative signal sequence is give in bold. A possible glycosaminoglycan attachment site is underlined. The first 40 amino-acid section of the mature protein is collagen-like, whilst the remainder of the sequence resembles keratin. The protein is
 10 glycine-rich and contains several repeats of the motif (C/S)1-4(Y/F), which is also found in structural proteins from insect egg shells. The tyrosines may be involved in cross-linking by formation of dityrosine-bridges by phenoloxidases. A similar protein is encoded by clone I (see Figure 8).

15 Figure 8 is a partial cDNA-sequence and cDNA-inferred polypeptide sequence of clone I. The inferred protein is glycine- and tyrosine-rich and resembles a cement protein of the reef-building polychaete *Pragmatopoma californica* (a component of the quinone-tanned cement in the tubes built by these marine worms).

20 Figure 9 is a DNA alignment between the protein sequence shown in Figure 8 and a cement protein from the polychaete *Pragmatopoma californica*.

Figure 10A shows a PAGE gel showing the proteins expressed from *E. coli* cells transformed with a truncated coding sequence from clone 64.

25

Figure 10B shows a Western blot corresponding to Figure 10A.

Figure 11.a and 11.b show alignments of the truncated clone 64 protein with various natural proteins.

30

Figures 12.a and 12.b show histological sections of hamster skin stained with various conventional stains.

Figures 13.a, 13.b and 13.c show immunoperoxidase-stained sections of hamster skin.

EXAMPLES

5

Ticks

Ticks were reared according to Jones *et al.*, (1988). All three developmental stages of *Rhipicephalus appendiculatus* were fed on Dunkin Hartley guinea pigs. When not
10 feeding, all ticks were maintained at 21 - 26°C and at 85% relative humidity.

Example 1 : Identification of proteins

Cement cones were collected from ticks (nymphs and adults) feeding on guinea pigs at
15 different points of the attachment period. The cones were homogenised in phosphate-buffered saline (PBS), to extract soluble proteins, and in hot alkali and acid to extract less soluble components (Kemp *et al.*, 1982; Jaworski *et al.*, 1992).

Protein patterns were analysed using SDS-PAGE; the resulting gel is displayed as
20 Figure 1A. Bands or spots corresponding with early expressed (group A) proteins and later expressed (group B) proteins were excised from the gel and used for the production of polyclonal antiserum. Good antisera could be obtained even against the less antigenic proteins provided that these proteins were not allowed to renature completely.

25

The resulting antisera were used to screen cDNA libraries and also for immunoblotting (see Figure 1B) and immunohistochemistry.

Proteins for which no good antiserum could be raised were blotted onto
30 polyvinylidene difluoride membranes for amino-terminal sequence determination. For amino acid sequencing, samples were run on an Applied Biosystems 494A "Procise sequencer" (Perkin-Elmer, Applied Biosystems Division, Warrington U.K.).

Electroblotted samples are run using Applied Biosystems "Mini-Blott" cartridge in the place of the standard cartridge. Bands of interest are excised from the membrane and cut into 1 x 3mm pieces for insertion into the cartridge. These are sequenced using the manufacturer's recommended programme for membrane-bound samples (Schagger and von Jagow, 1987; Matsudaira, 1987).

This information was then used to design oligonucleotides to screen a tick cDNA library for clones of interest.

10 Example 2 : cDNA library construction

Salivary glands were excised from 20 male and 20 female adult *R. appendiculatus* specimens that had been feeding on guinea pigs for two days. The glands were collected on dry ice in an Eppendorf tube. Messenger RNA was isolated using the FastTrack mRNA isolation kit (Invitrogen).

To synthesize cDNA and insert the cDNA into the Lambda Zap II vector, the ZAP cDNA synthesis kit (Stratagene) was used. Prior to unidirectional insertion of the cDNA into the lambda vector, the nucleic acid was fractionated over a Sephacryl S-400 column (Pharmacia).

A library (termed d2-I) was constructed from low molecular weight cDNAs (ranging from approximately 100 to 2,000 base pairs). The higher molecular weight fraction was used to construct a second library (d2-II). Packaging was performed using Packagene (Promega) packaging extracts. Approximately 1.5×10^6 plaque-forming units (PFU) of each library were amplified in XL-1-Blue cells (Stratagene) for subsequent use.

Example 3 : Screening of the d2-II cDNA library

Phagemids were excised *in vivo* from a randomly selected fraction of the library, and used to generate double-stranded pBluescript SK(-) plasmids in XL1-Blue cells (Stratagene), as described by Short *et al.*, (1988).

XL1-Blue colonies were plated out on ampicillin-containing LB (Luria-Bertani) agar plates, supplemented with 5-bromo-4-chloro-3-indolyl- β -D-galactopyranoside (X-Gal, Melford Laboratories, UK) and isopropyl- β -D-thiogalactopyranoside (IPTG, Novabiochem) for blue/white colony selection.

About 75 plasmids (from white colonies) with inserts ranging from 250 to 1000 base pairs (as determined by digestion with *PvuII* and electrophoresis over a 1% agarose gel) were selected for sequencing. The oligonucleotides used for sequencing correspond to the T3 and T7 primer sites in the pBluescript plasmid DNA.

Example 4 : Sequencing

Plasmids were purified from overnight cultures according to Goode and Feinstein (1992), alkali-denatured (Mierendorf and Pfeffer, 1987), and sequenced using the Sanger dideoxy-mediated chain termination reaction (Sanger and Coulson, 1975). Sequence data were analysed using the GCG sequence analysis software (Program Manual for the Wisconsin Package, 1994).

Protein database searches were performed at the National Centre for Biotechnology Information (NCBI) using the BLAST network service.

A number of clones were sequenced or partially sequenced. The sequences or partial sequences of those clones are shown in Figures 2 to 8 attached hereto. Explanations of the structure and features of the cloned sequences are given in the Brief description of the Figures above and in other parts of the description.

Example 5 : Production of the Cema Antiserum

An antiserum was produced against a prominent salivary gland protein of around 17kDa size.

5

For production of the polyclonal antiserum, salivary glands taken at day 6 of the adult feeding stage were homogenised in phosphate-buffered saline (PBS) and submitted to centrifugation (3 min; 12,000 x g). The proteins in the supernatant [i.e. the salivary gland extract (SGE)] were resolved over a 15% SDS-polyacrylamide gel, according to Laemmli (1970).

The gel was stained in an ice-cold 100 mM KCl solution and the 17 kDa protein was excised. The polyacrylamide section containing the protein was dried under vacuum, homogenised in PBS, mixed with an equal volume of Montanide ISA 50 adjuvant (Seppic, France) and subcutaneously injected into Dunkin Hartley guinea pigs. This procedure was repeated every 10 days. Serum was collected 10 days after the 4th injection.

Western blotting, performed according to Kyhse-Anderson (1984), showed a strong reaction of the antiserum with a 17kDa protein from the surface of cement cones (see Figure 1A). This protein, termed Cema, was present at all feeding stages of the ticks and could be easily obtained by washing of the cement cones in PBS.

Example 6 : Immunohistochemistry/ western blotting/ northern blotting/ *in situ* hybridisation

The polyclonal antisera were used in western blots and in staining sections of cement cones and salivary glands, taken at different stages of the feeding period. Where light-microscopy did not provide sufficient resolution to visualise the stain, electron microscopy was used.

For proteins against which no antisera could be produced, northern blots were instead performed using digoxigenin-labelled DNA probes constructed by random primer labelling (Sambrook *et al.*, 1989) using purified insert from the original clones. An anti-digoxigenin antiserum conjugated with alkaline-phosphatase allows probe
5 detection. In fact, *in situ* hybridisation may be more suitable for localising and following the expression of genes in the salivary glands, since immunohistochemistry when performed on salivary glands often results in a high background.

In conjunction with the SDS-PAGE data, these techniques allow determination of the
10 times at which specific proteins were expressed during the feeding period, where in the salivary glands they were produced and to which layer of the cone they contribute (thus better defining group A and B proteins).

**Example 7 : Construction of genomic libraries and examination of differential
15 expression.**

In order to evaluate the differential expression of tissue cement protein genes, the following procedure is presently being followed. Genomic tick DNA, digested with suitable endonucleases is inserted into the Lambda Fix II vector (Stratagene), which
20 allows for easy restriction mapping. Digoxigenin-labelled cDNA probes are used for library screening.

Regions flanking the coding sequences and introns are being sequenced and examined for the presence of sites that might play a role in the ordered expression of the proteins
25 (for example ecdysteroid or heat-shock response elements). It was thought that comparison of genes of proteins expressed simultaneously might reveal common upstream or downstream sequences that are responsible for the regulation of gene expression. All group A protein genes, for example, may have identical recognition sites for regulatory factors.

30

Prospective regulatory regions are coupled to a reporter gene, for example luciferase, transfected into suitable cells (*e.g. Drosophila* cells), and submitted to functional

assays. Gel retardation, DNA protection or band-shift assays are also being performed to confirm the existence of functional regulatory domains.

It is possible that a single promoter region controls the simultaneous expression of a whole series of genes. These genes are then most probably localised in close proximity to one another on the genome. To investigate this possibility, the digoxigenin-labelled probes are being used to localise the genes on the genome by means of *in situ* hybridisation, Southern blotting and genomic library screening.

10 Expression of cement proteins may be regulated by one or more haemolymph-borne factor(s). This possibility is being investigated by incubating salivary glands taken from animals that have only just attached to their host in tissue culture medium containing haemolymph from ticks that have been feeding for 24 or 48 hours. Blotting (northern and western) or reverse transcriptase-PCR is then used to determine whether
15 genes have been switched on or off.

Where there is evidence for a haemolymph-borne factor controlling the expression of cement genes, transplantation of salivary glands from ticks early in the feeding stage, to animals later in the feeding stage is being carried out, in case *in vitro* incubation
20 experiments do not provide adequately clear results. This factor has been identified by HPLC and other standard techniques.

Example 8 : Aggregation/ cross-linking studies - protein expression

25 Aggregation (polymerisation) and cross-linking are being determined using proteins extracted from salivary glands and cement cones, and also using expressed recombinant proteins. Cement cone proteins are being examined directly by protein hydrolysis followed by detection of di- or tri-tyrosines among the amino-acids. Other assays are also being used for the detection of cross-linking enzymes and SDS-PAGE
30 in the presence and absence of reducing agents to reveal inter- or intramolecular disulphide-bridges.

In addition, native group A and group B proteins are isolated from cement and salivary gland extracts by means of immunoprecipitation. This is only possible for the more soluble proteins. The molecular weight of the precipitated proteins is being determined by gel filtration. Comparisons are being made to the molecular weight of monomers as
5 determined by SDS-PAGE and western blotting, or as calculated from the cDNA derived protein sequences.

Other components co-precipitating with a given (A or B) protein are being identified by screening the cDNA-library with oligonucleotide probes designed using N-terminal
10 amino acid sequences.

The nature of eventual intermolecular bonds are also being determined. Hydrogen-bonds can be destroyed with urea and hydrophobic interactions with detergents. Detection of disulphide-bridges and cross-linked amino-acids can be performed as
15 described previously (Creighton, 1989; Malencik *et al.*, 1996).

Studying native proteins is not always straightforward. For example, it can be difficult to extract enough protein to enable exhaustive study. Also, non-specific protein interactions may occur. Many of these problems can be solved by the use of
20 recombinant proteins in their place.

Recombinant group A and group B proteins are therefore being expressed in bacteria or, where glycosylation and other post-translational modifications are crucial, in a eukaryotic (baculoviral) system. Those proteins which are not very soluble can be
25 expressed in fusion with thioredoxin using, for example, the ThioFusion system (Invitrogen). In this system, the proteins are provided with oligohistidine-tags, allowing easy purification by means of nickel-agarose chromatography (Janknecht *et al.*, 1991). Enterokinase and/or thrombin sites are incorporated to remove tags and fused proteins from the cement protein after purification, in cases where this is
30 necessary. In case aggregation of a protein takes place, or if interaction occurs between, for example, two different group A proteins, the nature of the bonds must be determined, as for the native proteins - see above. Expressed proteins which can be

recognised at all times by their histidine tags, or by the antisera were incubated with salivary gland extracts, in the absence and presence of specific and non-specific enzyme-inhibitors, in order to identify cross-linking.

5 The histidine-tagged expressed proteins can then be coupled to nickel-agarose and used in affinity-chromatography (Bugge *et al.*, 1992; Lu *et al.*, 1993) to isolate interacting proteins from salivary gland extracts or cement cones. These proteins can then be identified through the screening of cDNA libraries. By deletion and mutation experiments, domains or residues participating in the aggregation or cross-linking of
10 recombinant proteins, are thus being identified.

Example 9 : Expression of truncated cement protein (64TRP) in bacteria

Oligonucleotides with appropriate restriction enzyme sites were designed to permit
15 PCR cloning of an N-terminal fragment of clone 64, as shown in Figure 7. This fragment, known as 64P (amino acids 34 to 85) from the cDNA, was PCR cloned in-frame into the pET23 vector (Novagen). The construct, 64TRP (encoding amino acids 34 to 85), tagged onto the 6 x His-Tag of the pET23 vector, was obtained using standard PCR cloning methods (Sambrook *et al.*, 1989). The plasmid was transformed
20 into *E. coli* AD494 cells (Sambrook *et al.*, 1989).

The expressed histidine-tagged expressed 64TRP (truncated cement protein) was purified by means of nickel-agarose affinity chromatography (Janknecht *et al.*, 1991). The protein was analysed by SDS-PAGE (Leammli, 1970) and the resulting gel is
25 shown in Figure 10A. The expected protein band of approximately 7.8 kDa in size was obtained plus an extra upper band of about 10 kDa. The apparent size of the upper band indicates that it is not a dimer. Amino-terminal sequence analysis (Schagger and von Jagow, 1987; Matsudaira, 1987) of the two bands is currently being performed to determine the relationship between the two bands. A corresponding Western blot is
30 shown in Figure 10B.

Example 10: Expression of full-length cement protein (64P) in insect cells using recombinant baculovirus

The full length clone of the cement protein 64P (i.e. 144 amino acids in length) was
5 amplified from the cDNA using oligonucleotides containing appropriate restriction
enzyme sites, inserted into the C129.1His baculovirus vector and transfected into
Sodoptera fugiperda insect cells for eukaryotic expression (O'Reilly *et al.*, 1994).

Example 11: Similarity of the amino acid sequence of the *R. appendiculatus* cement 10 protein to sequences of other structural proteins

Protein database searches were performed at the National Centre for Biotechnology
Information (NCBI) using the BLAST for the amino acid sequences of both the
truncated and full length clone of the cement protein. Figures 11a and 11b show the
15 relationships between the 64P, the truncated 64TRP and other structural proteins.

The first 40 amino acids of the cement protein are strongly collagen-like (Fig. 11b)
and the rest of the sequence resembles keratin (Fig. 11a). The protein is glycine-rich
and contains several repeats of the motif (C/S) 1-4 (Y/F) resembling structural proteins
20 from *Drosophila melanogaster* (cuticular protein) and other insect egg shells, as well
as vertebrate cytokeratins including mammalian keratin complex 2 basic protein,
mouse keratin, human keratin, collagen type IV alpha and IPIB2 precursor. It would
seem that the cement protein has been designed to resemble the skin proteins of the
host, with the most likely aim of avoiding rejection of skin-tick attachment by the
25 host's natural immune defence mechanisms. The compositional resemblance of 64P
with its surrounding tissues may also facilitate the intimate binding between the
cement cone and the surrounding skin tissues (see Examples 12 and 13).

Example 12 : Histological studies

30

Histological studies using either haematoxylin and eosin or van Gieson stains
(Bancroft and Stevens, 1990) were performed on normal hamster skin sections and on

hamster skin sections on which *R. appendiculatus* ticks had been fed. Figure 12.a.A shows a normal skin section with intact epidermal, dermal and subcutaneous layers. The haematoxylin and eosin, and van Gieson stains indicate the expected tissue-types (i.e. basement membranes, collagen fibers and reticular fibers.) In skin sections on 5 which *R. appendiculatus* had fed, the tick cement cone is clearly attached to the skin (Figure 12.b.A) and can be seen embedded in the dermis (Figure 12.a.B and 12.b.B) with the mouth parts *in situ*. Comparatively, the cement cone seems to resemble normal skin basement membranes, collagen/reticulin fibers, in tissue stain uptake. Also, there is no evidence of infiltration by inflammatory cells in relationship to the 10 cement cone. The apparent absence of inflammation adds credence to the hypothesis that the structure and composition of the cement cone resembles that of skin tissues and is designed to avoid provoking a host response against the tick. The layered structure of the cement cone is clearly distinguishable in Figure 12.a.B. The apparent layering is probably due to the discontinuous deposition of cement proteins as the tick 15 alternates between salivation (i.e. secretion) and imbibing of host fluids.

Example 13: Immunohistochemical studies/Western blotting

The denatured recombinant 64TRP protein was further purified by cation-ion 20 exchange chromatography according to the manufacturer's recommendations (Pharmacia). The purified protein was used to raise polyclonal antiserum by subcutaneous injections of equal volumes of protein and Montanide ISA 50 adjuvant into Dunkin Hartley guinea pigs.

25 Western blotting (Kyhse-Anderson (1984)) showed a strong reaction of the antiserum to the denatured recombinant 64TRP cement protein bands (Figure 11B). Western blots with *R. appendiculatus* salivary gland and cement cone extracts from male and female ticks (prepared as previously described in Example 5) showed no reaction with the antiserum. The most probable explanation for the negative results is that the anti- 30 64TRP antiserum binding epitope(s) were not readily available on the extracted proteins owing to the conformation of the proteins.

Using the anti-64TRP antiserum, immunohistochemical studies were performed on sections of both normal hamster skin, and *R. appendiculatus* salivary glands, cement cones and cement cones attached to hamster skin on which *R. appendiculatus* ticks had been attached (Coligan *et al.*, 1991).

5

Using normal skin, the anti-64TRP antiserum reacted strongly with the basement membranes, epidermal and dermal tissues as well as the hair follicular structures (compare Figure 13.a.A with the control serum, Figure 13.a.B). In skin from hamsters on which ticks had been fed, sections of individual cement cones reacted with the antiserum (Figures 13.b. and Figures 13.c.). The reactions were mainly with the outermost layer and inner layer attached to the basement membrane of the skin. The reaction pattern with cement cones may indicate that 64P is a cement protein that lines the cement cone, possibly acting as a glue that binds the cement cone to the surrounding epidermal and dermal tissues.

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CLAIMS

1. A tissue cement protein having the amino acid sequence shown in Figure 3 or Figure 7 or containing any one of the partial amino acid sequences shown in any one of Figures 2, 4 to 6 and 8, related tissue cement proteins from blood-feeding parasites, preferably ticks, and functional equivalents thereof.
2. The tissue cement protein of claim 1 that is of the group A subtype as hereinbefore defined.
3. The tissue cement protein of claim 1 that is of the group B subtype as hereinbefore defined.
4. The tissue cement protein of any one of the preceding claims that binds to vertebrate tissues.
5. The tissue cement protein of any one of the preceding claims that is derived from a blood-feeding ectoparasite.
6. The tissue cement protein of claim 5 that is derived from ticks.
7. The tissue cement protein of claim 6 that is derived from Ixodid ticks.
8. The tissue cement protein of claim 7 that is derived from *Rhipicephalus appendiculatus*.
9. The tissue cement protein of any one of the preceding claims that is associated with one or more carbohydrate moieties.
10. The tissue cement protein of claim 9 that is associated with one or more glycosaminoglycan moieties.

11. The tissue cement protein of any one of the preceding claims that is expressed in recombinant form.
12. The tissue cement protein of any one of the preceding claims that is
5 associated with one or more peptides or polypeptides.
13. The tissue cement protein according to claim 12 which is associated with one or more self molecules to form a homodimer, homotrimer or homotetramer unit.
- 10 14. The tissue cement protein according to claim 12 associated with one or more non-self molecules to form a heterodimer, heterotrimer or heterotetramer unit.
15. The tissue cement protein of any one of the preceding claims that has
15 been genetically or chemically fused to one or more peptides or polypeptides.
16. The tissue cement protein of any one of the preceding claims that has been cross-linked to one or more peptides or polypeptides.
- 20 17. The tissue cement protein of any one of the preceding claims attached to a label.
18. The tissue cement protein of any one of the preceding claims attached to a toxin.
- 25 19. The tissue cement protein of any one of the preceding claims that is bound to a support, such as a resin.
20. A pharmaceutical composition comprising a mixture of group A and
30 group B tissue cement proteins according to any preceding claim in the absence of other parasite saliva proteins but in the presence of one or more compounds capable of cross-linking said tissue cement proteins, in conjunction with a pharmaceutically-

acceptable excipient.

21. A pharmaceutical composition comprising a mixture of group A and group B tissue cement proteins according to any one of claims 1 to 19 in the absence of other parasite saliva proteins in conjunction with a pharmaceutically-acceptable excipient.

22. A pharmaceutical composition comprising a mixture of group A and group B tissue cement proteins according to any one of claims 1 to 19 together with those saliva proteins necessary for the cement-hardening process, but in the absence of other parasite saliva proteins, in conjunction with a pharmaceutically-acceptable excipient.

23. The tissue cement protein according to any one of claims 1 to 19 or a pharmaceutical composition of any one of claims 20 to 22 for use in therapy.

24. Use of the tissue cement protein of any one of claims 1 to 19 as a pharmaceutical.

25. The tissue cement protein of any one of claims 1 to 19 for use as a vaccine or as a component of a vaccine.

26. Use of the tissue cement protein of any one of claims 1 to 19 as a vaccine or vaccine component.

25

27. A vaccine comprising the tissue cement protein of any one of claims 1 to 19.

28. A method of production of the vaccine of claim 27 comprising immunising an animal with the tissue cement protein of any one of claims 1 to 19.

30

29. A method of bonding animal tissue comprising bringing an animal tissue into conjunction with one or more tissue cement proteins of any one of claims 1 to 19 that are capable of forming a hardened cement.
- 5 30. The tissue cement protein of any one of claims 1 to 19 or a pharmaceutical composition of any one of claims 20 to 22 for use in the temporary or permanent bonding of tissues.
31. The tissue cement protein of any one of claims 1 to 19 for use as a
10 protective immunogen in the control of diseases caused by infections transmitted by arthropod parasites.
32. A nucleic acid molecule which encodes a tissue cement protein of any one of claims 1 to 19 or which hybridises with said nucleic acid molecule under
15 standard hybridisation conditions.
33. The nucleic acid molecule of claim 32 which comprises DNA, cDNA or RNA.
- 20 34. The nucleic acid molecule of claim 32 or claim 33 which comprises DNA.
35. A cloning or expression vector comprising a nucleic acid molecule of any one of claims 32 to 34.
- 25 36. The vector of claim 35 which is virus based.
37. The vector of claim 36 which is baculovirus based.
- 30 38. A host cell transformed or transfected with the vector of any one of claims 35 to 37.

39

39. A transgenic animal that has been transformed by the nucleic acid molecule of any one of claims 31 to 34 or a vector of one of claims 35 to 38.

40. A method of preparing a tissue cement protein of any one of claims 1 to 5 19, comprising expressing a vector according to any one of claims 35 to 37 in a host cell and culturing said host cell under conditions where said protein is expressed, and recovering said protein thus expressed.

FIG. 1B

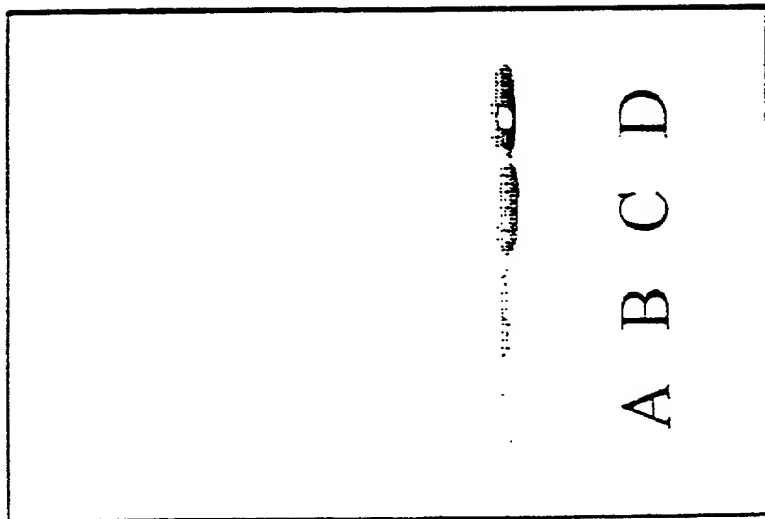
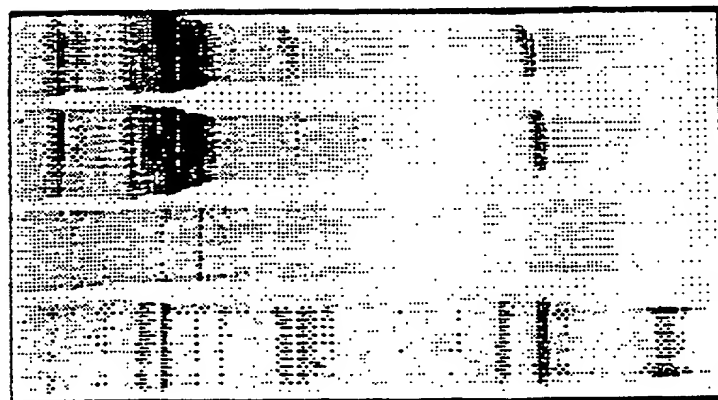


FIG. 1A



A B C D

FIG. 2

1 AAACCAAGGCAGGACACAGCAGCCATGAAGGCCCTTCGTTGCAGTCGCCCTTTGTCTGCA 60
M K A F V A V A L L S A

61 GTTCCCGTGGCACATGCTGCCCTCAAGACTGACGTAGCCAGTGGACCTGCCGGTCTGGT 120
V S V A H A A L K T D V A S G P A G S G

121 GCACTAAGTCTAGGAGTTGGAGGCTTCCCGTCCGGTGCTTCGCTTGGCAGCCTTAGTGCC 180
A L S L G V G G F P S G A S L G S L S G

181 GTAACCCCTCTGTGGTGGCTCTTCCGTGTCTGGCCGCCCTGGATCCCCCTGGATCGGCT 240
V T L S G A G S S V S G R P G S P G S A

241 GGTCCCTAGCTCTGGACCCGCGAGTGTCG 267
G P S S G P A V S

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FIG. 3

1 CGGACGCACAC'TCCTGCAGGAAGGTCATCTAGTTCGGCCAAACATGAAGCTGCTCTGTGCA 60
M K L L C A

61 CTAGCCCTCGTTGCCCTTGACTTCACATTCGGCAGCGCTTACCTTGGTGGCTTCGGCGGC 120
L A L V A L G L P F G S A Y L G G F G G

121 CTCGGTGGTTGGGTGGCGGTCTCGGTGCCATCTTTGGCCCAAGAGCTTATCCCGGTTTC 180
L G G W G G G L G A I F G P G A Y P G F

181 TATGGCCTTAACAGCGTGCACCTCTTTGGCGGCAAGTTCACCATCTCTTCGGGCGATTC 240
Y G L N S V H L L G G R F H L F G R F

241 CCGCCACCAACCGGTATTGGAGCTGCTGAAGCGCAGGGGAACCTAAGCCCATACCCCTCTT 300
P P P G I G A A E A Q G N L S P Y P L

301 GACATCAACACCGTCCAAGACCCGAACCTGGCCACCCCATGTACGCGTTGTCTACGGCGG 360
D I N T V Q D P N W P P H G T R C L R R

361 AGTCTTGGGGAGCGCCTCTGACCCCTGACCCAGTCCCAATTCCACAGGATGTGCTGTCCC 420
S L A G A P L T L T S P N S T G C A C P

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421 AGTCCCATTCAGTGCCCCAGCCATACCCAGTCCCACACCCACGACAAGTTCATACCCC 480
S P H S S A P A I P S P T P T S S I P

481 AGTGCCCTAGTCCCTACCCCGTCCCAATCCACAGTAACACCGAAGTTCACAAGACCGACGT 540
S A

541 CGTCGCCGCTACTCCAGGAGGACCAGTCCCTGCTCGGAGTCGGTGTACCCGGTGTCTCAGGCC 600
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601 AGGCGAACCAAGGGTCGTGGCCCTAAGCTTGATCCCAATAGAAAGTCATAACAATTTAGTCA 660

661 GTGAGCTCCACGTAATTTATGCATTACAATAAAGAAAAGTTTGTCTGGCAGTAAAAAAA 720

721 AAAAAAAAAA 730

FIG. 3(CONTD.)

FIG. 4

1 GATCGGCACGAGGTCAAGGGAGGCCCTCCTTCAGCAACAACAAGCATCGCAGGTTAAGGGA 60
D R H E V K G A L L Q Q Q A S Q V K G

61 GCCCTCAAGGGAGCAATCAAGGGTGTCTTCTTCAGCAACAAGCCCCAATCCCCAAGTCCAA 120
A L K G A I K G G L L Q Q Q A Q S Q V Q

121 GGAGCTCTTAAGGGAGCCGTCAAGGGAGCCCTCCTTCAGCAACAACAAGGCATCACAGGTC 180
G A L K G A V K G A L L Q Q Q Q A S Q V

181 AAGGGAGCCCTCAAGGGAGCCCATCAAGGTCTGTCTCCTTCATCAGCAAGCCCCAATCCCAA 240
K G A L K G A I K V C L L H Q Q A Q S Q

241 TCCCAAGTTCAGGGAGCTCTTAAGGGAGCTG 271
S Q V Q G A L K G A

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FIG. 5

1 GGAAGTAGCGAGCATCCGCACTGGGGTCTTTTGGCTGCATTGCTTTTCTTCTTCAGC 60
E V A S I R T G V F L A A F A F L L S A

61 GATCCATAACAATGGCCAGTCATGTAGATGCAGCCCCCACTCGACGTCCTATGCCATC 120
I H N N G Q S C V D A A P T R R P M P S

121 TCCTCCTGGATGTGCTGGTCCTGGCTGTTTACTGGTATGCTACTCTTCTAAGACCTGG 180
P P G C A G P G C F T G I A T L L R P G

181 TCAAGGACAGCAACCTGGTCAAGGACAGCAACCTGGTCAAGGGCGTCTCCAATGCCACG 240
Q G Q Q P G Q G Q Q P G Q G R P P M P R

241 TCCAGGACCTGTTCCAGGAACATCTGGATCACCTCAAGGAAGACCCCAATGGAGCACCTCG 300
P G P V P G T S G S P Q G R P N G A P R

301 TCCAGGACCTGTTCCCTGGAACATCTGGATCACCTCAAGGAAGACCTAAGCAAGACCTCG 360
P G P V P G T S G S P Q G R P N A R P R

361 TCCAGGACCTGTTCCCTGGAACACCAACTGTATCCTCTCCCGGATCATCTCCTGGTCATC 420
P G P V P G T P T V S S P G S S P G S S

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421 TCCAGGAATATCTCTAGGAACGCCCTCTAGGAACACCTCTAGGAACACCTCAAGGATCACC 480
P G I S L G T P L G T P L G T P Q G S P

541 TTTTGGATCATCTCTTGGATCATCGATAGGATCACCTCCTGCAACATCTCCTGGATCATC 540
F G S S L G S S I G S P P A T S P G S S

601 TTCTCCGTCACCTCCTGGATCAGCGAATGTGAACCTGCTGGTCTCGACCAATTCGCGG 600
S P S P P G S A N V N L L G P R P I R G

661 TCCTGGAAGGCATTGACGGGACCAGTTCTGCTGTGTATTCTCTCCGTGCACAAATGAGGGAA 660
P G R H

721 GGCATTGATGGGACCAGTTCTGCTGTGTATTCTCTCCGTGCACAGTGAGGGAATCTATCAA 720

FIG. 5(CONTD.)

721 TAGTGCAATAA 731

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1 GGCTTCGGCAGCCCACTCAGCGGTTTCGGCAGCCCACTCAGCGGTTTCGGCAGCCCACTC 60
G F G S P L S G F G S P L S G F G S P L

61 AGCGGCTTCGGCAGCCCACTCAGCGGATTTCGGTAGCCCACTCAGCGGATTTCGGTAGCCCA 120
S G F G S P L S G F G S P L S G F G S P

121 CTCAGCGGATTTCGGTAGCCCACTTCGGCAGCTACGGTCCCCCTGTCCATGGGTCTCGGAGCC 180
L S G F G S P F G S Y G P L S M G L G A

181 CCCAGGAGATTCCCCGGCGACCTCCGCCCTCATCTCTGAGCCCCACCTCCCGCCTTCCCGTT 240
P R R F P G D L R L I S E P T S R L P V

241 AGCGATGCCGCTACACCGCTGTCTGTCAGCCCGTCACAAAGCGCAGTGGTCCACACCGAG 300
S D A V Y T A V V Q P V T S A V V H T E

301 GGTCGCCATGTCACCGGCCAAGTACAGGAACACGTTGCAATCTAAGCTTTTCTAACCGCA 360
G P H V T G Q V Q E H V A I

361 AGCTATATTACGACGGATTAGTCAACACAGTCATCTTAAGCAAAATGTATCTAAATAAAA 420

421 TTTATCTGCCT 431

FIG. 6

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FIG. 7

1 GGAGATCACCTGCTTGCAAGGACAAACGTCCTAACACAGCCGCAAAATGAAAGCTTCTT 60
M K A F F

61 CGTTCTTCCCTTCTTCAACCGCGCACTGACGAATGCAGCAAGGCTGGTCGTCTGG 120
V L S L L S T A A L T N A A R A G R L G

121 AAGCGACCTGGATACATTTGGAAGGGTACACGGTAACCTATATGCCGGCATCGAAAGAGC 180
S D L D T F G R V H G N L Y A G I E R A

181 TGGCCCTCGTGGATACCCAGGGCTTACCGCATCGATTGGAGGCGAAGTGGTGCACGACT 240
G P R G Y P G L T A S I G G E V G A R L

241 CGGTGGTCGTGCCGGTGTGGGAGTGAGCAGCTACGGCTATGGTTACCCCTTCATGGGGCTA 300
G G R A G G V G S S Y G Y G Y P S W G Y

301 TCCGTATGGTGGATACCGTGGATACCGTGGATACCGTGGATACCGTGGATACAGGG 360
P Y G G Y G G Y G G Y G G Y D Q G

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361	TTTGGCTCTGCATACGGCGGCTACCCGGCTACTATGGCTACTACTATCCCAGTGGCTA	420
	F G S A Y G G Y P G Y Y G Y Y Y P S G Y	
421	CGGTGGGGCTACGGTGGTAGCTACGGTGGCAGCTACGGTGGTAGCTACACCTATCCCAA	480
	G G Y G G S Y G G S Y G G S Y T Y P N	
481	CGTTCGGGCTTCAGCTGGTGCCGCAGCTTGAGCTTCTCCTTCAGCGTCACAGTAAGAAAT	540
	V R A S A G A A A *	
541	CATGGAGCACCCGATCGAGAAATACAGAGGTTCTCAAAAGCGTACGGGATGCCAACCAGC	600
601	AAGAAATTGCGCCGCAAAATGTTGAGAACAAATACAAGTTTCTGTAAAAA	656

FIG. 7(CONTD.)

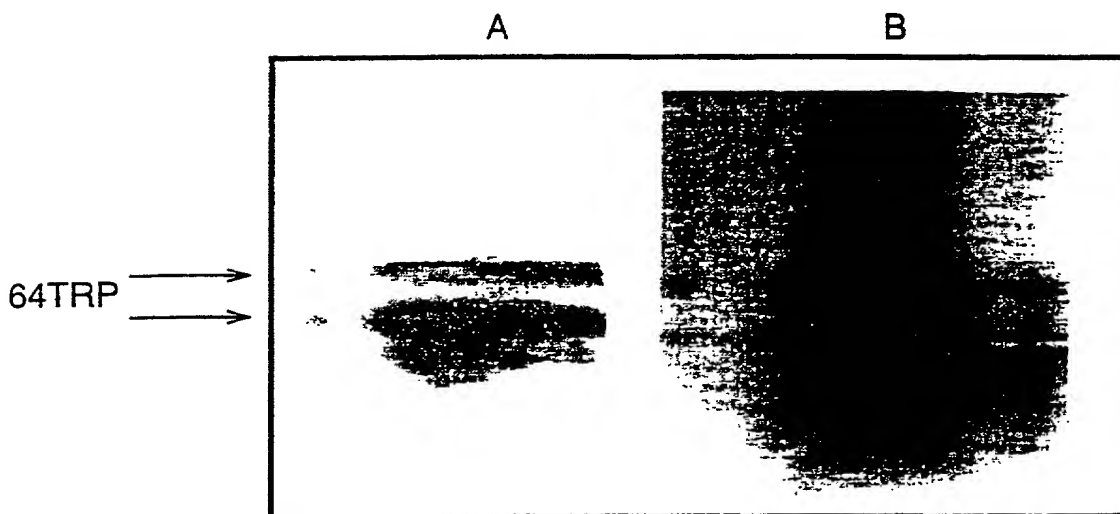
1	ACGGACTAGGTTTCGCTGGCGTCCCTCTTATTGGCGGATACGGCTACGGTCCTTTTCGTAG	60
	G L G F A G V P L I G G Y G Y G P F V G	
61	GAGCCTTCGCGTACGGCTTGTTGGGGTGCCCTCGGTGGCTATGGCTACCCCTTCGGAC	120
	A F A Y G L W G G L G G Y G Y P A F G L	
121	TCTCCTGGGTTCCACATGGTTTGGAGGCTTTGGAGCTTCTCCGCTGCTGCTGTTTCC	180
	S W V P H G F G G F G A S P S A A G F R	
181	GCTCGCTTTGGAGCCCTCTT	199
	S L W S L	

R. appendiculatus
GLGEAGVPLIGGYGYGPFVGAFAAYGLWGGLGGYGYPAEGLSWPHGEGFGASP
I.I: I.. :IIIIII: :I::II: :IIII.I: |::II:II::
GYGY.GAKKVGGYGYGAKLGGYGYG..AKIGGYGYGAKSGIQV.RALGGYGAGA

P.californica

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FIG. 10
EXPRESSION OF RECOMBINANT (TRUNCATED)
STRUCTURAL PROTEIN, 64TRP IN *Escherichia coli* CELLS



SDS-PAGE: (A) AND (B) WESTERN BLOT (USING ANTI-64TRP ANTISERUM) AND COOMASSIE BLUE STAINED 15% POLYACRYLAMIDE GEL, RESPECTIVELY OF IPTG-INDUCED *E.coli* CELLS EXPRESSING 64TRP PROTEIN. SAMPLES WERE SOLUBULISED AT 100°C IN SDS PRIOR TO LOADING THE GELS.

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FIG. 11a cDNA inferred sequence alignment between the protein sequence of 64P and other related structural proteins.

ker2	T	E	L	R	R	T	V	Q	G	L	E	I	E	L	Q	S	Q	L	A	L	K	Q	S	L	E	A	S	L	A	E	T	E	G	R	Y	C	V	Q	L	S	Q	I	Q	405
ker3	T	E	L	R	R	T	V	Q	G	L	E	I	E	L	Q	S	Q	L	A	L	K	Q	S	L	E	A	S	L	A	E	T	E	G	R	Y	C	V	Q	L	S	Q	I	Q	406
ker1															E	V	V	K	K	Q	C	I	G	V	Q	D	S	I	A	D	A	E	Q	H	G	E	H	A	I	K	D	A	R	29
ker4	S	E	L	N	R	V	I	Q	R	L	R	S	E	I	D	N	V	K	K	Q	I	S	N	L	Q	Q	S	I	S	D	A	E	Q	R	G	E	N	A	L	K	D	A	K	440
64																																										0		
ker2	S	Q	I	S	A	L	E	E	Q	L	Q	Q	I	R	A	E	T	E	C	Q	N	A	E	Y	Q	Q	L	L	D	I	K	T	R	L	E	N	E	I	Q	T	Y	R	S	448
ker3	S	Q	I	S	A	L	E	E	Q	L	Q	Q	I	R	A	E	T	E	C	Q	N	A	E	Y	Q	Q	L	L	D	I	K	T	R	L	E	N	E	I	Q	T	Y	R	S	449
ker1	G	K	L	T	D	L	E	E	A	L	Q	Q	C	R	E	D	L	A	R	L	L	R	D	Y	Q	E	L	M	N	T	K	L	S	L	D	V	E	I	A	T	Y	R	K	72
ker4	N	K	L	N	D	L	E	D	A	L	Q	Q	A	K	E	D	L	A	R	L	L	R	D	Y	Q	E	L	M	N	T	K	L	S	L	D	V	E	I	A	T	Y	R	T	483
64																																										31		
ker2	L	L	E	G	E	G	S	S	S	G	G																																466	
ker3	L	L	E	G	E	G	S	S	S	G	G																																467	
ker1	L	L	E	G	E	E	C	R	M	S	G	D	F	S	D	N	V	S	V	S	I	T	S	S	T	I	S	S	S	M	A	S	K	T	G	F	G	S	G	G	Q	S	115	
ker4	L	L	E	G	E	E	S	R	M	S	G	E	C	A	P	N	V	S	V	S																							518	
64																																										36		
ker2	S	G	G	S	Y	G	G	S	S	S	G	G	S	Y	G	G	S	S	G	G	S	Y	G	G	S	S	G	G	S	Y	G	G	S	Y	G	G	S	Y	G	G	S	S	506	
ker3	S	G	G	S	Y	G	G	S	S	S	G	G	S	Y	G	G	S	S	G	G	S	Y	G	G	S	S	G	G	S	Y	G	G	S	Y	G	G	S	Y	G	G	S	S	507	
ker1	G	G	R	G	S	Y	G	G	R	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G	157	
ker4	G	G	G	G	Y																																						553	
64	N	L	Y	A	G	I	E	R	A	G	P	R	G	Y	P	G	L	T	A	S	I	G	G	E	V	G	A	R	L	G	G	R	A	G	V	G	V	S	S	Y	G	Y	78	

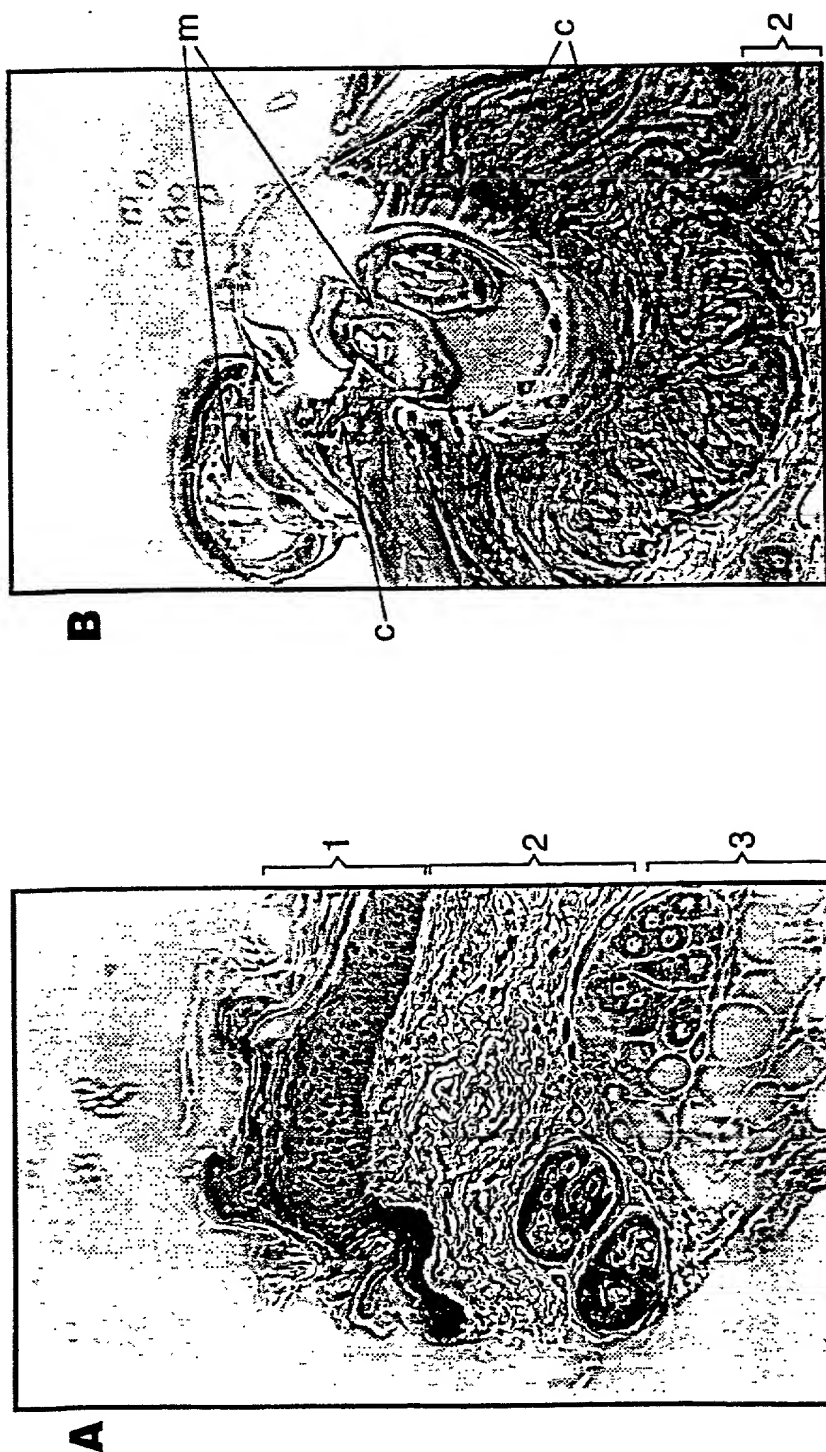
Sequences are: ker1=keratin complex2 basic protein, ker2=mouse keratin, ker3=mouse epidermalK protein, ker4=human keratin protein, 64P=R. appendiculatus putative structural protein

FIG. 11b cDNA inferred sequence alignment between the protein sequences of 64TRP and other structural proteins.

[illegible]

Sequences are: coll=Glycine-rich GX protein-motif LGG, col2=collagen type IV alpha protein, col3=IPB2 precursor protein, col4=Glycine rich cell wall protein, 64TRP=R. appendiculatus truncated structural protein

FIG. 12a HISTOLOGICAL STUDIES OF THIN HAMSTER SKIN SECTIONS STAINED WITH HAEMATOXYLIN AND EOSIN

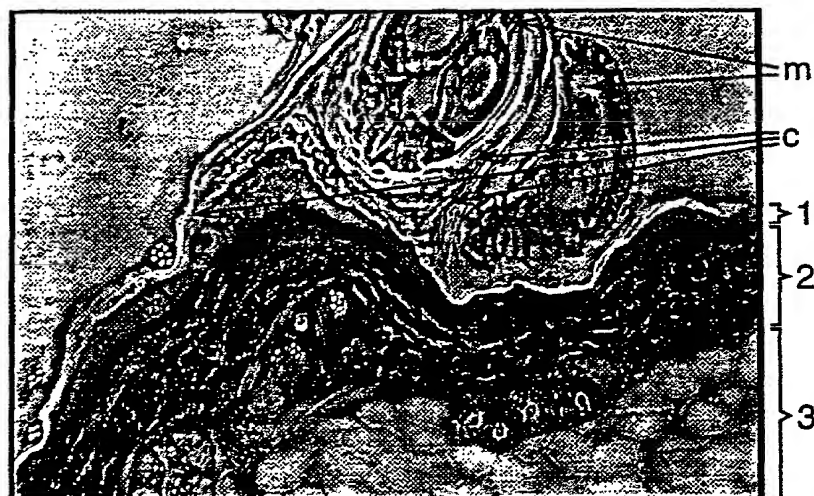


(A) AND (B) = HISTOLOGICAL SECTIONS OF HAMSTER SKIN STAINED WITH HAEMATOXYLIN AND EOSIN;
 (A)=NORMAL SKIN SECTION SHOWING EPIDERMIS-1, DERMIS-2 AND SUBCUTANEOUS LAYER-3.
 (B)=SKIN SECTION SHOWING AN EMBEDDED TICK (*R. appendiculatus*) CEMENT CONE. STAINING: PINK=
 BASEMENT MEMBRANES/COLLAGEN FIBRES, UNSTAINED=RETICULIN FIBRES, BLUE=CELL
 NUCLEI. C REFERS TO CEMENT CONE AND M TICK MOUTH PARTS. ABOUT 20x

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FIG. 12b

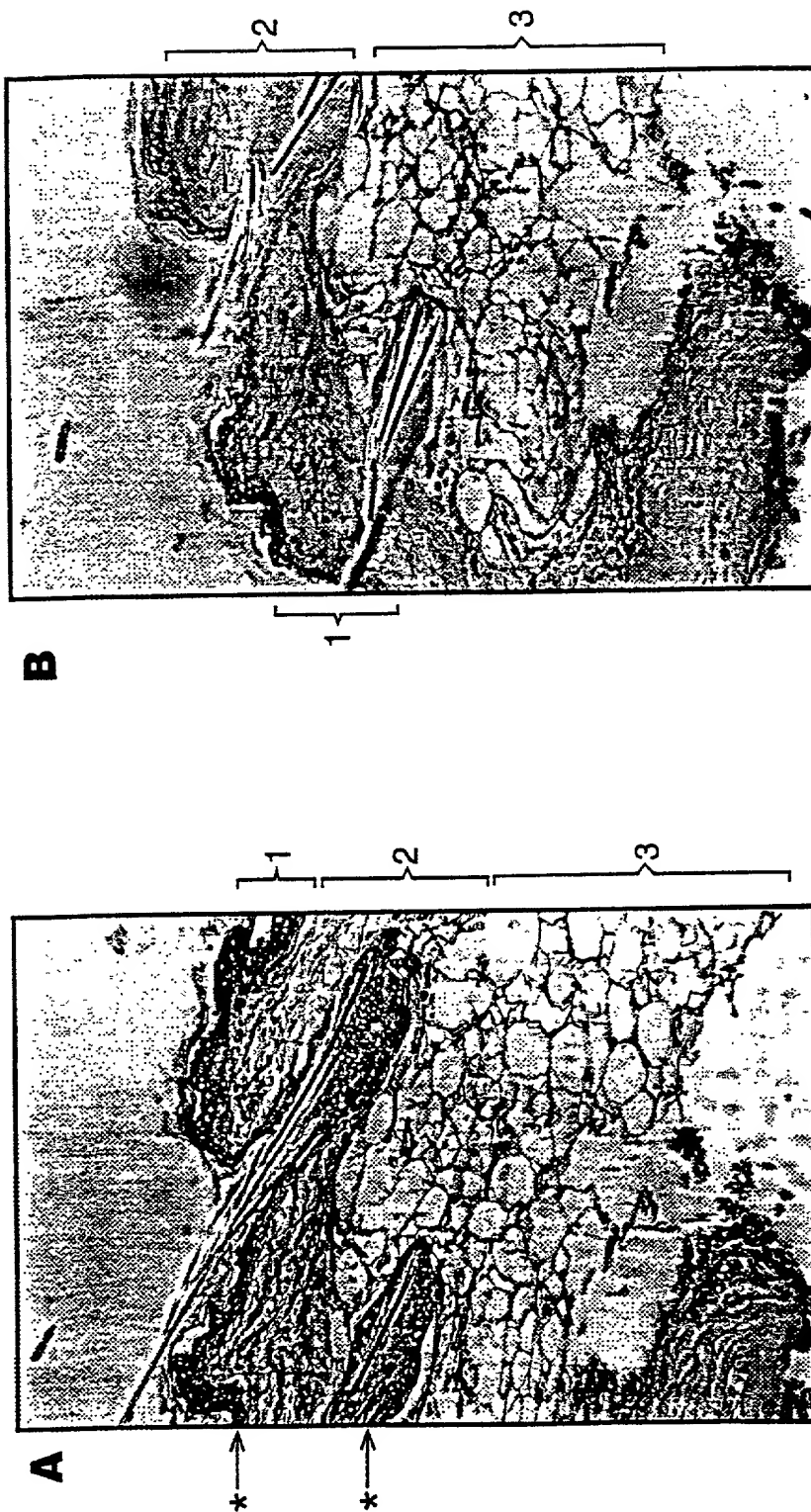
HISTOLOGICAL STUDIES OF THIN HAMSTER SKIN SECTIONS
STAINED WITH HAEMATOXYLIN AND EOSIN, AND VAN GIESON STAINS.

A**B**

(A) AND (B) = HISTOLOGICAL SECTIONS OF HAMSTER SKIN SHOWING ATTACHED TICK (*R. appendiculatus*) CEMENT CONES (i.e. POST FEEDING) STAINED WITH (A) HAEMATOXYLIN AND EOSIN STAIN (H & E), (B) VAN GIESON STAIN. 1=EPIDERMIS, 2=DERMIS AND 3=SUBCUTANEOUS LAYER. STAINS: H & E-PINK=BASEMENT MEMBRANES/COLLAGEN FIBRES, BLUE=CELL NUCLEI, VAN GEISON-RED=COLLAGEN FIBRES, YELLOW=BASEMENT MEMBRANES/RETICULIN AND ELASTIN FIBRES. C=CEMENT CONE AND M= TICK MOUTHPARTS. ABOUT 20x.

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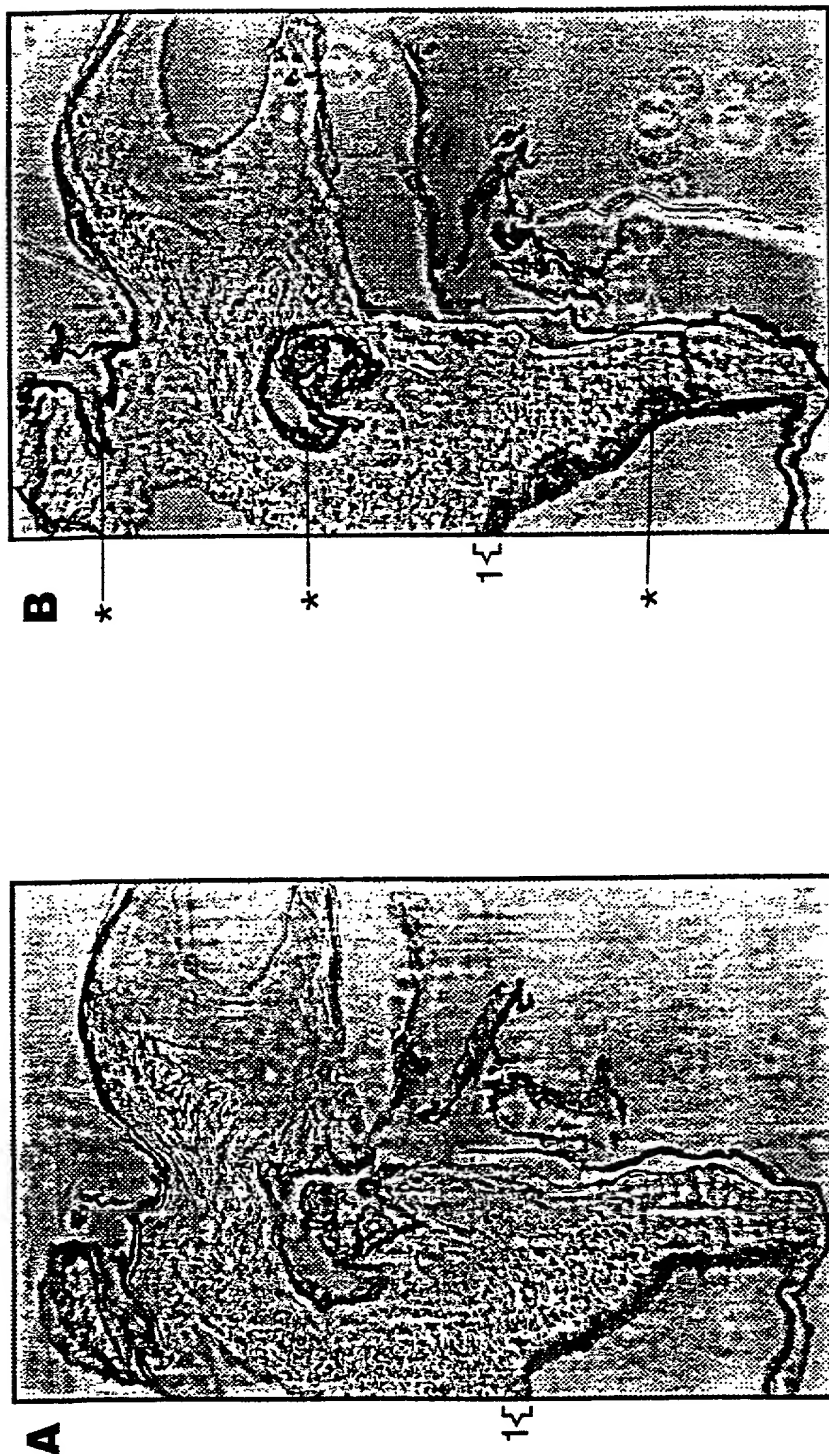
FIG. 13a IMMUNOPEROXIDASE STUDIES ON HAMSTER SKIN SECTIONS USING ANTI-64TRP ANTISERUM



(A) AND (B) = NORMAL HISTOLOGICAL SECTIONS OF HAMSTER SKIN INCUBATED WITH ANTI-64TRP ANTISERUM AND A POSITIVE CONTROL ANTISERUM RESPECTIVELY, AND DEVELOPED WITH PEROXIDASE LINKED SECONDARY ANTISERUM. 1= EPIDERMIS, 2=DERMIS AND 3= SUBCUTANEOUS LAYER; * = AREAS OF SKIN SHOWING POSITIVE REACTIONS TO ANTI 64TRP ANTISERUM. ABOUT 20X

18/19

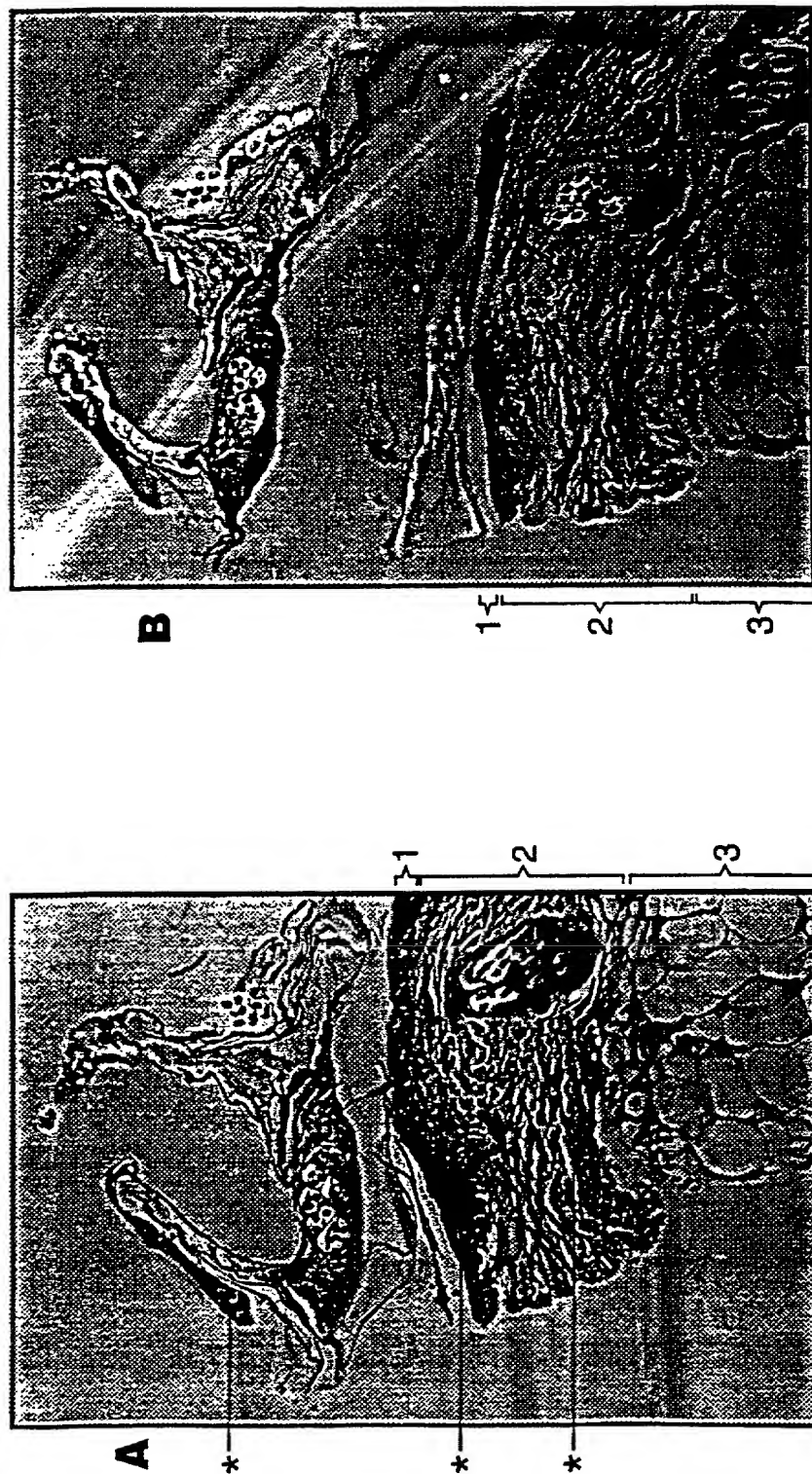
FIG. 13b IMMUNOPEROXIDASE STUDIES ON *R. appendiculatus* CEMENT CONE SECTIONS USING ANTI-64TRP ANTISERUM



(A) AND (B) = THIN SECTIONS OF *R. appendiculatus* CEMENT CONES INCUBATED WITH EITHER A POSITIVE CONTROL ANTI-SERUM OR ANTI-64 TRP ANTI-SERUM RESPECTIVELY, AND DEVELOPED WITH PEROXIDASE LINKED SECONDARY ANTISERUM. 1x=PART OF SKIN EPIDERMIS *==AREAS OF SKIN SHOWING POSITIVE REACTIONS TO ANTI-64TRP ANTI-SERUM. ABOUT 20x

19/19

FIG. 13c IMMUNOPEROXIDASE STUDIES USING ANTI-64TRP ANTISERUM ON THIN HAMSTER SKIN SECTIONS POST FEEDING WITH *R. appendiculatus*



(A) AND (B) = THIN SECTIONS OF HAMSTER SKIN POST-FEEDING WITH *R. appendiculatus* SHOWING A CEMENT CONE STILL ATTACHED, INCUBATED WITH ANTI-64TRP ANTISERUM AND A POSITIVE CONTROL ANTISERUM RESPECTIVELY, AND DEVELOPED WITH PEROXIDASE LINKED SECONDARY ANTISERUM. 1=EPIDERMIS, 2=DERMIS AND 3=SUBCUTANEOUS LAYER. * = AREAS OF SKIN SHOWING POSITIVE REACTIONS TO ANTI 64TRP ANTISERUM. ABOUT 20x

DECLARATION AND POWER OF ATTORNEY FOR PATENT APPLICATION

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below under my name.

I believe that I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled

TISSUE CEMENT

the Specification of which

☐ is attached hereto
☒ was filed on May 12, 2000
as Application Serial No. 09/554,547
and was amended on _____ (if applicable).

I hereby state that I have reviewed and understand the contents of the above-identified Specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, 1.56(a).

I hereby claim foreign priority benefits under Title 35, United States Code, §119 (a)-(d) or § 365(b) of any foreign application(s) for patent or inventor's certificate, or §365(a) of any PCT international application which designated at least one country other than the United States of America, listed below and have also identified below, by checking the box, any foreign application for patent or inventor's certificate, or of any PCT international application having a filing date before that of the application on which priority is claimed.

<u>APPLICATION</u> <u>NUMBER</u>	<u>PRIOR FOREIGN FILED APPLICATION(S)</u> <u>COUNTRY</u> <u>(MONTH/DAY/YYYY)</u>	<u>PRIORITY</u> <u>CLAIMED</u>
9723945.3	GB 12th November 1997	Yes

I hereby claim the benefit under Title 35, United States Code §119(e) of any United States provisional application(s) listed below.

APPLICATION NUMBER(S)

FILING DATE (MM/DD/YYYY)

I hereby claim the benefit under Title 35, United States Code, §120 of any United States application(s), or §365(c) of any PCT international application designating the United States of America, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT international application in the manner provided by the first paragraph of Title 35, United States Code §112, I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations §1.56 which became available between the filing date of the prior application and the national or PCT international filing date of this application.

<u>U.S. Parent</u> <u>Application No.</u>	<u>PCT Parent</u> <u>Number</u>	<u>Parent Filing</u> <u>(MM/DD/YYYY)</u>	<u>Parent Patent</u> <u>Number (if applicable)</u>
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PCT/GB98/03397 12th November 1998

The undersigned hereby authorizes the U.S. attorney or agent named herein to accept and follow instructions from Carpmaels & Ransford as to any action to be taken in the Patent and Trademark Office regarding this application without direct communication between the U.S. attorney or agent and the undersigned. In the event of a change in the persons from whom instructions may be taken, the U.S. attorney or agent named herein will be so notified by the undersigned.

I hereby appoint as my attorneys or agents the following persons: Jack Matalon, (Attorney, Registration No. 22,441); Stefan J. Klauber (Attorney, Registration No. 22,604); David A. Jackson (Attorney, Registration No. 26,742); Michael D. Davis (Attorney, Registration No. 39,161); William C. Coppola (Attorney, Registration No. 41,686); Christine E. Dietzel (Agent, Registration No. 37,309); Donald J. Cox (Attorney, Registration No. 37,804); and Michael A. Yamin (Agent, Registration No. 44,414), said attorneys or agents with full power of substitution and revocation to prosecute this application and transact all business in the Patent and Trademark Office connected therewith.

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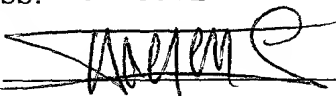
I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further, that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

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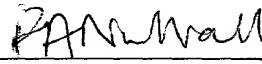
DATE 30/6/00

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DATE 30 June 2000